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**INSULTOS AMBIENTAIS E NUTRICIONAIS  
PROGRAMAM PARA A SÍNDROME METABÓLICA NA  
VIDA ADULTA**

MARINGÁ

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**INSULTOS AMBIENTAIS E NUTRICIONAIS  
PROGRAMAM PARA A SÍNDROME METABÓLICA NA  
VIDA ADULTA.**

Tese apresentada ao Programa de Pós-graduação em Ciências Biológicas (Área de concentração - Biologia Celular e Molecular) da Universidade Estadual de Maringá, para obtenção do grau de “Doutor em Ciências Biológicas”.

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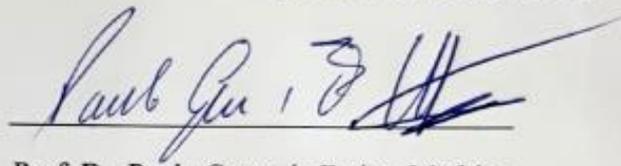
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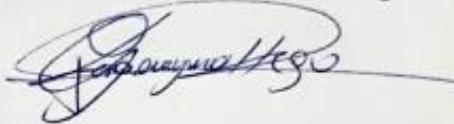
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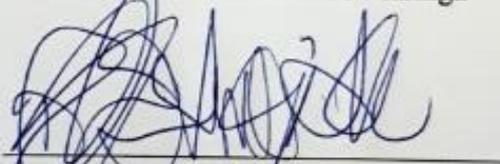
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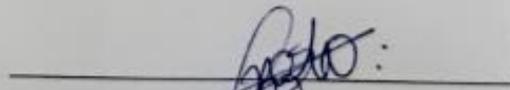
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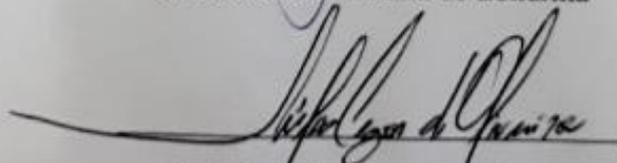
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## **BIOGRAFIA**

**Vander Silva Alves** nasceu na cidade de Araruna no PR, no dia 24 de Janeiro de 1987. É licenciado em Ciências Biológicas pela Faculdade Integrado de Campo Mourão (ano de 2008), Mestre em Ciências Biológicas (PBC/UEM) (ano de 2012) e atualmente realiza o Doutorado no Programa de Pós-graduação em Ciências Biológicas (PBC/UEM). Possui experiência em Biologia Celular e Fisiologia.

## APRESENTAÇÃO

Esta tese é composta de dois artigos científicos. O primeiro é intitulado “**Particulate matter from air associated to obesity enhances vulnerability to metabolic impairment and oxidative stress markers**”. O trabalho demonstra que a administração via gavagem de material particulado ( $PM_{<10}$ ), durante a lactação é um agente programador para o indivíduo, possuindo capacidade de exacerbar as consequências provocadas pela hipernutrição no início da vida e subsequente exposição a uma dieta hiperlipídica na vida adulta. Em consonância com as regras do Programa de Pós-graduação em Ciências Biológicas, o artigo foi redigido de acordo com as normas da revista científica *Environmental Science and Pollution Research* com atual fator de impacto 2,8. O segundo é intitulado “**High fat diet during adolescence in male rats is able to programs blood pressure increase and lead to others metabolic dysfunctions at adulthood**”. O trabalho demonstra que a exposição à dieta hiperlipídica durante a adolescência programa para o desenvolvimento da síndrome metabólica na vida adulta, evidenciando dessa forma que a adolescência, bem como a gestação e lactação, é também uma janela de programação. Em consonância com as regras do Programa de Pós-graduação em Ciências Biológicas, o artigo foi redigido de acordo com as normas da revista científica *Plos one* com atual fator de impacto 3,234.

## RESUMO GERAL

INTRODUÇÃO - A obesidade e a síndrome metabólica (MetS) são enormes desafios para países desenvolvidos e em desenvolvimento. A alta prevalência dessas doenças tem sido apontada como consequência de mudanças na composição da dieta, aumento da ingestão alimentar, baixos níveis de atividade física, alterações na microbiota intestinal e, mais recentemente, exposição à poluição do ar. Evidências mostram que a exposição a estressores ambientais durante períodos cruciais do desenvolvimento, como gestação, lactação e adolescência, pode levar ao desenvolvimento de distúrbios metabólicos na vida adulta. Essa ideia é conhecida como origens desenvolvimentistas da saúde e doença (DOHaD). Assim, evidências crescentes indicam que a exposição a estressores ambientais durante essas janelas vulneráveis contribuem para a patogênese não transmissível, como obesidade, doença coronariana, diabetes mellitus tipo 2 (DMT2) e alguns tipos de câncer. Além disso, tem sido demonstrado que o excesso de peso, induzido pela hipernutrição precoce durante a vida pós-natal, é um fator crucial para o desenvolvimento da obesidade e disfunções cardiometabólicas. Neste contexto, os animais de ninhada reduzida (NR) têm sido amplamente estudados como um modelo importante para o desenvolvimento da programação metabólica e consequente desenvolvimento de MetS na prole. Os animais NR caracterizam-se pela presença de hiperfagia, resistência à leptina, resistência à insulina (RI) e aumento do estresse oxidativo. Outro modelo consistente para estudar a obesidade e a MetS pode ser a exposição à dieta rica em gordura (HFD). Existe uma relação entre a exposição à HFD durante os períodos de gestação e lactação e as alterações na pressão arterial na vida adulta. Também foi sugerido que a adolescência é uma fase suscetível de programação para a MetS. Por outro lado, um número crescente de evidências tem mostrado que o material particulado (PM) do ar desempenha um papel crucial no desenvolvimento de disfunções metabólicas e doenças crônicas, como a obesidade e o DMT2. Recentemente, dados do nosso grupo mostraram que a exposição materna à PM durante a vida perinatal foi capaz de levar a uma programação metabólica na prole na vida adulta. Embora os efeitos da PM sejam muito discutidos atualmente, as consequências de sua exposição por via oral durante a vida perinatal em modelos animais de obesidade são limitadas.

**OBJETIVOS** - **(a)**: Primeiro: verificar se a exposição materna de  $PM_{<10}$  durante a gestação e a lactação poderia exacerbar tanto as disfunções de curto como de longo prazo em prole de NR que ingeriram HFD na idade adulta. **(b)**: Segundo: verificar se a

exposição a HFD durante a adolescência pode levar a disfunções cardiometabólicas na vida adulta.

**MÉTODOS - (a):** Ratas prenhas foram alojadas em gaiolas individuais e randomizadas em dois grupos no dia 7 da gestação (GD). O primeiro grupo recebeu o veículo óleo de milho (OIL-Mothers) e o segundo recebeu por gavagem PM <10 (PM -Mothers) na dose de 50 µg de PM <10/rata. Durante todo o tratamento as mães receberam água *ad libitum* e alimentaram-se de dieta padrão para roedores (Nuvital®, Curitiba, Brasil). No dia pós-natal (PND) 1, todas as ninhadas foram ajustadas para 9 filhotes por mãe (com equilíbrio entre macho e fêmea). Para induzir a supernutrição precoce, no PND 3, os filhotes foram ajustados para 3 machos por mãe. Os filhotes das mães tratadas com óleo de milho foram denominados SL-OIL, enquanto aqueles das mães tratadas com PM<10 foram anotados como SL-PM. Avaliamos um lote da prole ao desmame, no PND 21 (n = 7 / grupo). Outro lote de ratos foi avaliado no PND 90 (n = 7/grupo). Também utilizamos outro lote de descendentes no PND 90 para avaliar o efeito combinado de HFD em ratos que foram expostos ao MP no início da vida (n = 7/grupo). Para isso, o SL-OIL e o SL-PM receberam por 30 dias (do PND 90 até o PND 120) HDF; Banha de 35%; 5,817 kcal/g). Avaliaram-se a composição do leite, homeostase da glicose (ipITT e ivGTT), perfil lipídico, um marcador de inflamação e alguns parâmetros de estresse oxidativo. **(b):** Ratos Wistar adolescentes (30 a 60 dias) foram expostos a dieta hiperlipídica (HFD, 35% de gordura). Os animais de controle tiveram acesso a ração comercial normal (NFD, 4,5% de gordura). A pressão arterial, frequência cardíaca e pressão de pulso foram verificadas em ratos com 120 dias de idade.

**RESULTADOS - (a):** A exposição de PM<10, coletada em área urbana de alta densidade de tráfego, durante a gestação e lactação levou a exacerbação das consequências a curto e longo prazo na superalimentação neonatal (induzida por ninhadas reduzidas) e subsequente ingestão de HFD na idade adulta. Neste estudo, mães tratadas com PM<10 durante a gestação e lactação apresentaram exacerbação das disfunções metabólicas causadas pela exposição à hipernutrição pós-natal precoce na prole, tanto no desmame quanto no final da vida. Além disso, PM<10 durante a vida perinatal foi capaz de agravar o fenótipo obeso na idade adulta após um segundo desafio com ingestão de HFD. **(b):** adolescentes com dieta hiperlipídica apresentaram alterações na ingestão calórica durante o período da dieta, bem como no ganho de peso, perfil lipídico e parâmetros cardiovasculares na vida adulta. Em conjunto, estes achados sugerem que a HFD durante

a adolescência, tem consequências a longo prazo na saúde e, desta forma, programa para o desenvolvimento da MetS mais tarde na vida.

**CONCLUSÃO - (a):** Exposição materna a  $PM_{<10}$  durante a gestação e lactação, foi capaz de exacerbar disfunções no metabolismo em modelos animais de obesidade, aumentando a intolerância à glicose e dislipidemia mais tarde. Entretanto, após um desafio nutricional com HFD na idade adulta, as disfunções metabólicas se tornam mais evidentes, demonstrando que a exposição ao MP no início da vida piora quando associada a insultos nutricionais, aumentando o estresse oxidativo, obesidade e levando à dislipidemia na idade adulta. **(b):** exposição à dieta hiperlipídica durante a adolescência levou a níveis mais elevados de pressão arterial sistólica e pressão arterial média na vida adulta. Além disso, a redução exacerbada da pressão arterial em resposta à injeção de hexametônio observada em animais com HFD sugere que o aumento da pressão arterial pela HFD durante a adolescência pode ser devido a uma maior atividade do sistema nervoso simpático.

## GENERAL ABSTRACT

1 **INTRODUCTION** – Obesity is an enormous challenge for developed and developing  
2 countries. This great prevalence have been pointed as consequence of changes in diet  
3 composition, increases in food intake, low levels of physical activity, changes in gut  
4 microbiome and more recently air pollution exposure. Several evidences have been  
5 shown that exposure to environmental stressors during crucial periods of developmental  
6 such as gestation, lactation and adolescence could lead to health or development of  
7 metabolic disorders in adulthood. This idea is known as developmental origins of health  
8 and disease (DOHaD) concept. Thus, increasing evidence has indicated that exposure to  
9 environmental endocrine and immune disruptors during these vulnerable windows  
10 contribute to the noncommunicable pathogenesis, such as obesity, coronary heart disease,  
11 type 2 diabetes mellitus (T2DM), and some types of cancer. It has been shown that  
12 overweight inducing by early overnutrition during postnatal life is a crucial factor for  
13 development of obesity and cardiometabolic dysfunctions, in this context, the small litter  
14 (SL) animals have been widely studied as an important model for development of  
15 metabolic programming and metabolic syndrome (MetS) in the offspring. SL animals are  
16 characterized by present hyperphagia, leptin resistance, insulin resistance (IR) and  
17 increase of oxidative stress. Other consistent model to study obesity and MetS may be  
18 exposure to high-fat diet (HFD). There is a negative relationship between exposure to  
19 HFD during the gestation and lactation periods and changes in blood pressure in adult  
20 life. It has also been suggested that adolescence is a susceptible phase for programming  
21 to MetS. On the other hand, a growing number of evidences have been shown that PM  
22 from air plays a crucial role in the development of metabolic dysfunctions and chronic  
23 disease such as obesity and T2DM. Recently data of our group shown that maternal PM  
24 exposure during perinatal life was able to lead a metabolic programing in the offspring at  
25 adulthood. Although the effects of PM are much discussed nowadays, the consequences  
26 of its exposure by the oral route during perinatal life in obesity animal models are limited.

27 **AIMS:** (a) - First: we hypothesized that maternal PM<sub><10</sub> exposure during pregnancy  
28 and lactation could exacerbate both short- and long-term dysfunctions on small litter  
29 offspring who fed HFD at adulthood. (b) – The second: our hypothesis argues that HFD  
30 during adolescence can lead to cardiometabolic dysfunctions at adult life.

31 **METHODS** - (a) : Pregnant rats were housed in individual cages and randomized into  
32 two groups on the gestation day (GD) 7. The first group received the vehicle corn oil

33 (OIL-Mothers) and second received by gavage PM<sub><10</sub> (PM-Mothers) at a dose of 50 µg  
34 of PM<sub><10</sub>, collected from a high-density-traffic urban area, during gestation and lactation.  
35 During all treatment mothers received water *ad libitum* and fed a balanced chow diet  
36 (Nuvital<sup>®</sup>, Curitiba, Brazil). On the post-natal day (PND) 1, all the litters were adjusted  
37 to 9 pups per dam (with balance between male and female). To induce early overnutrition,  
38 on the PND 3, the pups were adjusted to 3 male per dam. Offspring from the mothers  
39 treated with corn oil were denominated as SL-OIL, whereas those from the mothers  
40 treated with PM<sub><10</sub> were noted as SL-PM. We evaluated a batch of the offspring at  
41 weaning, on the PND 21 (n=7/group). Another batch of rats was evaluated at PND 90  
42 (n=7/group). We also used another batch of offspring at PND90 to evaluate the effect  
43 between HFD in rats that were exposure to PM in early life (n=7/group). For this, the SL-  
44 OIL and SL-PM received for 30 days (from PND 90 until PND 120) HFD; 35% lard;  
45 5.817 kcal/g). It was evaluated the milk composition, glucose homeostasis (ipITT and  
46 ivGTT), lipid profile, one inflammation marker and some oxidative stress parameters.  
47 **(b):** Adolescent Wistar rats (30 to 60 day-old) were exposed to a HFD (35% of fat).  
48 Control animals had access to normal commercial chow (NFD, 4.5% of fat). Blood  
49 pressure, heart rate and pulse pressure were verified in 120-day-old rats. Student t-test  
50 was used to compare groups.

51 **RESULTS - (a):** PM<sub><10</sub> exposure during pregnancy and lactation may exacerbate in  
52 offspring the short- and long-term consequences on neonatal overfeeding (induced by  
53 small litter) and subsequent ingestion of HFD at adulthood. In this study, mothers treated  
54 with PM<sub><10</sub> during critical periods of life (gestation and lactation) presented exacerbation  
55 of the metabolic dysfunctions caused by expose to early postnatal overnutrition in male  
56 offspring, both at early and later in life. Furthermore, PM<sub><10</sub> during perinatal life was able  
57 to aggravate the obese phenotype at adulthood after a second challenge with HFD  
58 ingestion. **(b):** HFD adolescents showed changes in calorie intake during the diet period,  
59 as well in the weight gain, lipid profile and cardiovascular parameters at adulthood. Taken  
60 together, these finding suggest that HFD during adolescence, have long term  
61 consequences on health and this way programs the development of cardiometabolic  
62 syndrome later in life.

63 **CONCLUSION – (a):** Maternal exposure to PM<sub><10</sub> was able to exacerbate dysfunctions  
64 on metabolism in obese animal model offspring, increasing glucose intolerance and  
65 dyslipidemia later in life. However, after a nutritional challenge with HFD at adulthood,  
66 metabolic dysfunctions become more evident, demonstrating that PM exposure in early

67 life worsens when combined with nutritional insults, increasing oxidative stress, obesity  
68 and leading to dyslipidemia at adulthood. **(b):** HFD exposition during adolescence  
69 programs to higher levels of systolic blood pressure and mean blood pressure later in life.  
70 In addition, the exacerbated blood pressure reduction in response to hexamethonium  
71 injection observed in HFD animals suggest that the increased blood pressure programed  
72 by HFD during adolescence may depends on sympathetic nervous system, which is an  
73 important predictor for cardiovascular death.

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101 **Particulate matter from air associated to obesity enhances vulnerability to metabolic**  
102 **impairment and oxidative stress markers in male rat offspring**

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118 Air pollution • Particulate matter • Perinatal life • Metabolic programming • Insulin  
119 resistance • Oxidative stress

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130 **Abstract**

131 **Background/Aims:** Several evidences have been shown that exposure to environmental  
132 stressors during crucial periods of development such as gestation, lactation and  
133 adolescence could lead to disease at adulthood. Furthermore, some reports have been  
134 shown that particulate matter (PM<sub><10</sub>) exposure combined with high fat diet (HFD) was  
135 able to exacerbate obesity potentiating chronic inflammation and insulin resistance in  
136 mice. We hypothesized that PM<sub><10</sub> exposure during pregnancy and lactation could  
137 exacerbate both short- and long-term dysfunctions on neonatal overfeeding rats.  
138 **Methods:** Samples of PM<sub><10</sub> from an urban area of Cotonou, Benin, were suspended in  
139 corn oil and administered by gavage (50 µg PM<sub><10</sub>/day) in pregnant and lactating rats  
140 (PM-Mothers group). The offspring were analyzed in three different ages: post-natal day  
141 (PND) 21, 90 and 120. **Results:** Our results showed that PM exposure lead to a several  
142 consequences in the offspring at weaning, where SL-PM at PND 21 showed an increase  
143 in body weight (bw) (p < 0.001), retroperitoneal (p < 0.05) and mesenteric (p < 0.001) fat  
144 pad stores and fasting glucose (p < 0.05). At PND 90 no statistical difference was found  
145 on the bw, fat pad stores, fasting glucose and fasting insulin. However, SL-PM showed  
146 an increased in total cholesterol (TC) (p < 0.01) and triglycerides (TG) (p < 0.05).  
147 Furthermore, during intravenous glucose tolerance test (ivGTT), an increase of glucose  
148 plasma levels (p < 0.01) was observed at SL-PM in the AUC, followed by a blood insulin  
149 level increment in the first 5 minutes at the test (p < 0.05). After HFD challenge, at PND  
150 120 the SL-PM showed higher weight gain (p < 0.01), increase of the retroperitoneal (p  
151 < 0.05) and mesenteric (p < 0.05) fat pad stores and higher insulin resistance (IR)  
152 compared to the SL-OIL. In addition, a significative reduction of the levels of glutathione  
153 (p < 0.05), glutathione transferase (p < 0.05) and superoxide dismutase (p < 0.005), was  
154 observed in the SL-PM compared to SL-OIL. **Conclusion:** Our data demonstrated that  
155 PM<sub><10</sub> exposure during pregnancy and lactation was able to exacerbate in the offspring  
156 metabolic disfunctions caused by exposure to early postnatal overnutrition, both at  
157 weaning and later in life, mainly after a second challenge with HFD ingestion. Thus, we  
158 suggest that places with poor air quality can lead to exacerbation of preexisting metabolic  
159 dysfunctions.

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168 **1. Introduction**

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170 Obesity is an important challenge for developed and developing countries, since  
171 promote a great impact on public health, and in this way, interfering on social and  
172 economic aspects from individuals and communities (Fruhbeck et al. 2013, Visscher et  
173 al. 2017). Reaching pandemic status, it has been estimated the quantity of overweight or  
174 obese among adults is 40% (Sung et al. 2019) and, this increased prevalence have been  
175 pointed as consequence of changes in diet composition, increases in food intake (Astrup  
176 &Brand-Miller 2012, Swinburn et al. 2009), low levels of physical activity (Hill &Peters  
177 2013), changes in gut microbiome (Tilg &Kaser 2011) and more recently ongoing air  
178 pollution exposure (Sun et al. 2009).

179 Several evidences have been shown that exposure to environmental factors and  
180 endocrine and immune disruptors during crucial periods of developmental such as  
181 gestation, lactation and adolescence could contribute to development of  
182 noncommunicable pathogenesis, such as obesity, coronary heart disease, type 2 diabetes  
183 mellitus (T2DM), and some types of cancer at adulthood (de Oliveira et al. 2013,  
184 Grandjean et al. 2015). This idea is known as developmental origins of health and disease  
185 (DOHaD) concept (de Oliveira et al. 2013, Grandjean et al. 2015, Reynolds et al. 2015,  
186 Vickers 2014).

187 It has been shown that overweight induced by early overnutrition during postnatal  
188 life is a crucial factor for development of obesity and cardiometabolic dysfunctions, in  
189 this context, the small litter (SL) animals have been widely studied as an important model  
190 for development of metabolic programming and metabolic syndrome (MetS) in the  
191 offspring (Schmidt et al. 2001, Stefanidis &Spencer 2012). Other consistent model to  
192 study obesity and MetS may be exposure to high-fat diet (HFD). This diet is known by  
193 to lead an increase fat pad deposition and gain in body weight, both in humans and  
194 experimental (Halima et al. 2018).

195 Particulate matter (PM) from air plays a crucial role in the development of these  
196 dysfunctions (Brook et al. 2017, Rajagopalan &Brook 2012). The mechanisms by which  
197 the PM leads to chronic disease may be related to tissue or systemic inflammation and  
198 oxidative stress (Brook et al. 2017, Laing et al. 2010, Rajagopalan &Brook 2012, Rao et

199 al. 2015). Indeed, some reports have shown that PM exposure combined with HFD was  
200 able to exacerbate obesity by potentiate chronic inflammation and insulin resistance in  
201 mice, evidencing a relationship between HFD and PM (Sun et al. 2005, Sun et al. 2009).

202         Recently, data from our group showed that maternal PM exposure during  
203 pregnancy and lactation was able on leadin metabolic programing in the rat offspring at  
204 adulthood (Miranda et al. 2018). Furthermore, it has been defended that organism with  
205 preexisting disease, such as MetS are more sensitive to PM exposure, than which may  
206 exacerbate their symptoms besides increase their risk for hospitalization and chronic  
207 disease (Clementi et al. 2019). Although the effects of PM are much discussed nowadays,  
208 the consequences of its exposure by the oral route during perinatal life associated to  
209 obesity at adulthood are limited.

210         Thus, we hypothesized that maternal PM <10 exposure during pregnancy and  
211 lactation in rats lead to metabolic programming in the offspring and it could exacerbate  
212 both short- and long-term dysfunctions, such as increased body fat deposition, glycemia,  
213 insulinemia and lipid profile on SL offspring who fed HFD at adulthood.

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## 226 **2. Material and Methods**

### 227 *Animals and experimental groups*

228 The experiments and results were reported in accordance with the ARRIVE  
229 guidelines for experiments involving animals (Kilkenny et al. 2010). All experimental  
230 protocols were approved by the Ethics Committee in Animal Research (CEUA n°  
231 5681231014) and conducted at the Laboratory of Secretion Cell Biology of the State  
232 University of Maringá.

233 Adult male and female Wistar rats, from the Central Animal House at the State  
234 University of Maringá, Paraná, Brazil, were housed in the Animal House of the  
235 Department of Biotechnology, Genetics and Cellular Biology in polypropylene cages (45  
236 cm/30 cm/15 cm) under light controlled conditions with a 12h light-dark cycle (07:00  
237 a.m. to 07:00 p.m.) and a temperature of  $22.0 \pm 2$  °C. After, the animals were mated at a  
238 proportion of three females to each male. We daily verified changes in the oestrous cycle  
239 by vaginal smears and pregnancy was confirmed by the presence of sperm. Pregnant rats  
240 were housed in individual cages and randomized into two groups on the gestation day  
241 (GD) 7. By gavage, both, the first group received the vehicle corn oil (OIL-Mothers) and  
242 second received PM<sub><10</sub> (PM-Mothers) at a dose of 50 µg of PM<sub><10</sub>, once a day, during  
243 gestation and lactation. During all treatment mothers received water *ad libitum* and were  
244 fed with a balanced chow diet (Nuvital®, Curitiba, Brazil).

245 We used 7 dams/group to obtain the litters and for this study, we choose randomly  
246 one offspring/group/litter. On the post-natal day (PND) 1, all the litters were adjusted to  
247 9 pups per dam (with balance between male and female). To induce early overnutrition,  
248 on the PND 3, the pups were adjusted to 3 male per dam. Offspring from mothers treated  
249 with corn oil were denominated as SL-OIL, whereas those from mothers treated with  
250 PM<sub><10</sub> were designated as SL-PM (Figure 1).

251 We evaluated a group of offspring at weaning, on the PND 21 (n=7/group).  
252 Another group of rats was evaluated at PND 90 (n=7/group). We also used another batch  
253 of offspring at PND90 to evaluate the effect between HFD in rats that were exposure to  
254 PM in early life (n=7/group). For this, the SL-OIL and SL-PM received for 30 days (from  
255 PND 90 until PND 120) HDF (35% lard; 5.817 kcal/g) produced basing on Barella et al.  
256 (2012) (Figure 1).

257

### 258 *Particulate matter sampling*

259 Particulate matter <10  $\mu\text{m}$  ( $\text{PM}_{<10}$ ) samples were collected in an urban area of  
260 Cotonou (Benin) and characterized for physicochemical composition at Dunkerque  
261 (France) as previously described Cachon et al. The PM is a mix of compounds that  
262 includes inorganic and organic components, such as Al, Fe, Mg, Mn, Na, Pb, Zn and  
263 carbonaceous materials; volatile organic compounds such benzene, toluene, ethylbenzene  
264 and o-xylene; polycyclic aromatic hydrocarbons, mainly fluoranthene, pyrene, chrysene,  
265 benzo [a] pyrene; paraffins such as tricosane, tetracosane, heptacosane, tritriacontane; and  
266 fatty acids such as hexadecanoic acid, octadecanoic acid, oleic acid and tetradecanoic acid  
267 (Cachon et al. 2014).

268

#### 269 *PM<sub><10</sub> reconstitution*

270 The  $\text{PM}_{<10}$  stock solution was prepared by reconstitution with 3 mL of corn oil  
271 and was sonicated for 30 minutes (Sonic Dismembrator Model 100, Fischer Scientific,  
272 Waltham, MA, USA). The gavage solutions were individually prepared using a stock  
273 solution (Miranda et al. 2018).

274

#### 275 *Dose solution administration*

276 The treatment started at the gestation day (GD) 7 and the female rats who received  
277 50  $\mu\text{g}$  of  $\text{PM}_{<10}$  solution daily corresponding to 200  $\mu\text{g}$   $\text{PM}_{<10}/\text{kg}$  of body weight (bw)  
278 were called PM-Mothers. All rats weighed approximately 250g of bw, so we administered  
279 a constant quantity of 250  $\mu\text{L}$  of reconstituted  $\text{PM}_{<10}$  per day at a dose of 50  $\mu\text{g}$  of  $\text{PM}_{<10}$ .  
280 This dose was constant, mimicking the daily exposure. The calculation for dose  
281 administration was performed on a baseline basis on the low dose of diesel oral exposure  
282 as described previously by Danielsen et al. (2008). The control group, called OIL-  
283 Mothers, received a corn oil solution at a constant volume of 1 mL/kg of bw. These  
284 solutions were administered by gavage from GD 7 to PND 21 (from 8:00 am to 9:00  
285 a.m.).

286

#### 287 *Milk collection*

288 Milk samples were collected at PND 21 before 2 hours pups had been weaned  
289 from their dams. Dams were anaesthetized with thiopental (0.2 mL, ip), and so to induce  
290 milk secretion, a synthetic oxytocin, Oxiton (5 U.I./mL, União Química S/A, São Paulo,  
291 Brazil), was injected (0.5 mL ip). Milk was collected in a sterile Pasteur pipette by

292 manually massaging the nipple as described previously (DePeters & Hovey 2009). Milk  
293 samples (1 mL/dam) were stored at -80° C for subsequent analysis.

294

#### 295 *Intraperitoneal insulin tolerance test (ipITT)*

296 After 6-hour fast the offspring at 120 days old (n = 6), was submitted to ipITT.  
297 After that, they received an injection of insulin (1 U/kg of body weight), and blood  
298 samples were collected, as previously reported (de Oliveira et al. 2011). Blood samples  
299 were obtained from the tail vein at 0, 15, 45 and 60 min after injection. Glucose levels  
300 were measured using the Accu-Chek Aviva system (Roche Diagnostics). The constant for  
301 the insulin tolerance test (Kitt) was calculated using formula  $Kitt (\%/min) = 0.693/t_{1/2}$ ,  
302 where  $t_{1/2}$  was calculated from the slope of the plasma glucose concentration during  
303 ipITT (Bonora et al. 1989).

304

#### 305 *Intravenous glucose tolerance test (ivGTT)*

306 At PND 90 and PND 120 the offspring (n=6 for each group and age) underwent  
307 surgery consisting of silicone cannula implantation into the right jugular vein as previously  
308 described (de Oliveira et al. 2011). After 12-h fasting, a glucose load (1 g/kg bw) was  
309 injected into the cannula. Blood samples (400 µl) were collected immediately prior to  
310 glucose injection (0 min) and then at 5, 15, 30 and 45 min, and the obtained plasma samples  
311 were stored at -20 °C for further analysis. To maintain blood volume, a corresponding  
312 volume of saline (0.9%) was infused through the cannula.

313

#### 314 *Biochemical analysis*

315 The blood glucose concentration was determined using the glucose oxidase method  
316 (Trinder 1969) with a commercial kit (Gold Analisa<sup>®</sup>, Belo Horizonte, MG, Brazil). The  
317 triglyceride (TG), total cholesterol (TC) and high-density lipoprotein (HDL) levels were  
318 measured by colorimetric commercial kits (Gold Analisa<sup>®</sup>, Belo Horizonte, MG, Brazil).  
319 Low-density lipoprotein (LDL) and very low-density lipoproteins (VLDL) levels were  
320 found using the Friedewald formula (Friedewald et al. 1972).

321 The insulin levels of plasma were determined using a radioimmunoassay (Scott et  
322 al. 1981) with a gamma counter (Wizard<sup>2</sup> Automatic Gamma Counter, TM-2470,  
323 PerkinElmer<sup>®</sup>, Shelton, CT, USA). For the radioimmunoassay, human insulin was used as  
324 a standard, and detection was performed using an anti-rat insulin antibody (Sigma-Aldrich<sup>®</sup>,

325 St. Louis, MO, USA) and <sup>125</sup>I-labelled recombinant human insulin (PerkinElmer<sup>®</sup>, Shelton,  
326 CT, USA). The intra- and inter-assay coefficients of variation were 12.2% and 9.8%,  
327 respectively, for insulin. The detection limit for the insulin levels was 1.033 pmol/l.  
328 Homeostasis model assessment: insulin resistance (HOMA-IR) and β-cell function  
329 (HOMA-β), were determined by formulas as previously described (Matthews et al. 1985).

### 330 *Fat pad stores assessment*

331 The animals were euthanized by decapitation method and fat pad stores was  
332 removed and weighed, to measure fat tissue accumulation.

### 333 *Sample collection to oxidative stress and inflammation profile*

334 The liver of the animals was collected, weighed, washed in Sodium Phosphate  
335 Buffer (PBS 0.1 M, pH 7.4) and stored in a freezer at -80 °C until the date of the  
336 biochemical tests (n=6 each group and age). A portion of the liver was homogenized  
337 200 mM potassium phosphate buffer (pH 6.5), in a five-fold dilution and uses to  
338 determine all parameters (Borges et al. 2018).

339

### 340 *Lipid hydroperoxide (LOOH) levels and Reduced Glutathione (GSH) levels*

341 The GSH levels in the liver were determined according to the adapted method of  
342 Sedlak and Lindsay (Borges et al. 2018, Sedlak & Lindsay 1968), which 25 μL of  
343 homogenate from the sample plus 975 μL of 5,5'-dithiobis-2-nitrobenzoic acid were read  
344 at 412 nm. Individual values were interpolated based on a GSH standard curve and are  
345 expressed as μg of GSH/g of tissue. The other part of the homogenate was centrifuged for  
346 20 min at 9000 × g, and a part of supernatant was used to measured lipid hydroperoxide  
347 (LOOH). LOOH concentrations were determined using an extinction coefficient of 4.3  
348 mmolar 1/cm, and the results are expressed as mmol/mg of tissue. Readings were  
349 performed at 560 nm using a spectrophotometer as previously describe (Jiang et al. 1991).

350

### 351 *Catalase enzyme activity (CAT)*

352 Determination of CAT activity in the liver was performed using the method of  
353 Aebi (1984) with adaptations (Aebi 1984). In a 96-well plate containing 200 μl of a 20

354 mM solution (5 mM Tris/EDTA Buffer, pH 8.0, 30% hydrogen peroxide and distilled  
355 water) were added 10  $\mu$ l of the diluted sample, or in the case of white, 10  $\mu$ l of distilled  
356 water. The absorbance was then measured at 240 nm.

357

#### 358 *Superoxide dismutase (SOD) and Glutathione S- transferase (GST) enzyme activity*

359 The enzymatic assay for SOD is based on the ability of SOD to inhibit the  
360 autooxidation of pyrogallol (Marklund & Marklund 1974). Readings were done at 405 nm  
361 using a spectrophotometer. The results are expressed as U of SOD/mg of protein. The  
362 enzymatic activity of GST was determined according to the method of Warholm et al.  
363 (Warholm et al. 1985). The tests were performed at 340 nm using a spectrophotometer,  
364 with an extinction coefficient of 9.6 mmolar 1/cm. The results are expressed as  
365  $\mu$ mol/min/mg of protein.

366

#### 367 *Myeloperoxidase activity (MPO)*

368 The precipitate from liver homogenate centrifugation was resuspended in 80 mM  
369 potassium phosphate buffer that contained 0.5% hexadecyltrimethylammonium. The  
370 samples were homogenized and centrifuged for 20 min at 11,000  $\times$  g at 4  $^{\circ}$ C. The reaction  
371 was done in a 96-well plate using tetramethylbenzidine. Enzymatic activity of  
372 myeloperoxidase (MPO) was read at 620 nm using a spectrophotometer. The results are  
373 expressed as units of optical density (OD)/min/mg of protein (Borges et al. 2018).

374

#### 375 *Statistical analyses*

376 The data are presented as the mean  $\pm$  SEM and were subjected to Student's t test.  
377 A value of  $p < 0.05$  was considered statistically significant using GraphPad Prism version  
378 6.0 for Windows (GraphPad Software Inc., San Diego, CA, USA).

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### 382 3. Results

#### 383 *Impact of PM exposure in maternal health and milk composition*

384 As shown in Table 1, any changes were found in maternal weight during  
385 pregnancy and lactation and there was no difference in maternal fat pads. PM-Mothers  
386 had higher fasting glucose and lower fasting insulin (14.5%,  $p < 0.01$ ; 50%,  $p < 0.01$ ,  
387 respectively). In the plasma profile, we observed that PM-Mothers increased LDL (22%  
388  $p < 0.05$ ), while decreased HDL, HOMA-IR and HOMA- $\beta$  levels (32.8%,  $p < 0.0001$ ;  
389 40.3%,  $p < 0.05$ ; 68%,  $p < 0.0001$ , respectively). On the other hand, any difference was  
390 observed in the TC, VLDL and TG levels.

391 In breast milk, PM mothers displayed higher insulin and TG levels compared to  
392 SL-OIL (167%,  $p < 0.0001$ ; 22.1%,  $p < 0.05$ ), as showed in Table 2.

393

#### 394 *Effects of PM exposure in offspring biometric and biochemical parameters at PND 21*

395 SL-PM group did not present changes in birth weight (Fig. 2a), however, they  
396 were heavier (Fig. 2b) than SL-OIL ( $p < 0.05$ ) at weaning. Retroperitoneal (Fig. 2c) and  
397 mesenteric (Fig. 2e) fat pads were increased by 47% and 18% ( $p < 0.05$ ;  $p < 0.01$ ),  
398 respectively in SL-PM offspring in PND 21.

399 As shown in the Table 3, at PND 21, SL-PM presented increased fasting glucose  
400 (+11%,  $p < 0.05$ ), when compared with SL-OIL. In contrast, fasting insulin level was  
401 decreased in these animals (-55%,  $p < 0.01$ ). In SL-PM group HOMA-IR and HOMA- $\beta$   
402 were decreased (-45%,  $p < 0.01$ ; -49%,  $p < 0.001$ , respectively).

403 SL-PM group, at PND21, displayed decreased HDL (-14.5%,  $p < 0.005$ ) and  
404 increased VLDL and TG levels (+44% and +48%,  $p < 0.001$ , respectively). No statistical  
405 difference was observed in TC and LDL levels (Table 3).

406

#### 407 *Effects of PM dams exposure in pregnant and lactating on the biometric and biochemical* 408 *parameters in the offspring at PND 90*

409 As shown in the Table 3, at PND 90 no statistical difference was found on the bw,  
410 as well as in the fat pad stores between SL-PM and SL-OIL groups. The same was  
411 observed in fasting glucose and fasting insulin. On the other hand, at PND 90 the animals  
412 presented an increased at TC and TG (49%,  $p < 0.01$ ; 55%,  $p < 0.05$ , respectively) in SL-  
413 PM group compared to the SL-OIL group.

414 Glucose intolerance was detected in the SL-PM offspring at PND 90 (Fig. 3a).  
415 These animals showed an increase of glucose plasma levels during intravenous glucose  
416 tolerance test (ivGTT) with an increase of 10.5% ( $p < 0.001$ ) in the AUC compared to the  
417 SL-OIL offspring (Fig. 3b). Although, the SL-PM group, at PND 90 (Fig. 3c) showed a  
418 blood insulin level increment in the first 5 minutes on the ivGTT compared with the SL-  
419 OIL rats ( $p < 0.05$ ), the AUC remained unchanged between the groups (Fig. 3d).

420

421 *Effect of HFD on the body weight, food intake and glucose-insulin homeostasis in SL-PM*  
422 *rat offspring at PND 120.*

423 Although no difference was observed in food intake from PND 90 to PND 120  
424 (Fig. 4a and 4b), in the evolution of the bw, SL-PM showed a higher weight gain from  
425 PND 90 until PND 120 (Fig. 4c), represented by the AUC (Fig. 4d), the PM group was  
426 37.1% higher than in the SL-OIL group ( $p < 0.01$ ) (Fig. 4c and 4d).

427 In addition, as observed in the Table 4, after a HFD ingestion, from PND 90 to  
428 PND 120, the SL-PM group exhibited a 12.3% and 25.2% increase in the retroperitoneal  
429 and mesenteric fat pad stores compared to the OIL offspring ( $p < 0.05$ ). Despite no  
430 statistical difference was observed in plasmas levels of glucose and insulin between the  
431 animals at PND 120, the PM group presented an increase in the TC of 25.6% ( $p < 0.001$ ),  
432 but no statistical difference was found on TG levels between the groups (Table 4).

433 During the ivGTT, no difference was observed between the groups at PND 120  
434 (Fig. 5a and 5b). However, the SL-PM group, showed a blood insulin level increment  
435 (40%,  $p < 0.05$ ) in the first 5 minutes on the test compared with the SL-OIL, the AUC  
436 remained unchanged ( $p > 0.05$ ; Fig. 5c and 5d). During the insulin sensitivity assessment,  
437 detected by ipITT (Fig.5e), the glucose uptake in the SL-PM group was expressively  
438 decreased compared to the OIL offspring ( $p < 0.01$ ; Fig. 5f), and an increase was observed  
439 with HOMA-IR and HOMA- $\beta$  index (Table 4).

440 *Effects of PM exposure in pregnant and lactating dams on oxidative stress status in liver*  
441 *after HFD exposure of their offspring at PND 120*

442           In the assessment of the oxidative status of the liver from offspring at PND 120  
443 no difference was found in the levels of CAT between the groups. The same happened  
444 with the levels of LOOH and MPO of the liver. On the other hand, the PM group showed  
445 a significant reduction of the levels of GSH, GST and SOD ( $p < 0.05$ ), compared to SL-  
446 OIL offspring (Fig. 6).

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#### 464 **4. Discussion**

465 To the best of our knowledge, this is the first report to describe whether ambient  
466 PM<sub><10</sub> exposure during pregnancy and lactation may exacerbate in offspring the short-  
467 and long-term consequences on neonatal overfeeding induced by SL and subsequent  
468 ingestion of HFD at adulthood. In this study, male offspring rats from mothers treated  
469 with PM<sub><10</sub> during perinatal life (gestation and lactation) presented exacerbation of the  
470 metabolic dysfunctions caused by exposure to early postnatal overnutrition, both at early  
471 and later in life. Furthermore, PM<sub><10</sub> during perinatal life was able to aggravate the obese  
472 phenotype at adulthood after a second challenge with HFD ingestion.

473 Ambient air pollution has been implicated with adverse pregnancy outcomes such  
474 as mortality in children and adults (Buka et al. 2006). In the present investigation, PM<sub><10</sub>  
475 exposure during gestation and lactation did not alter bw from dams. On the other hand,  
476 an increase of fasting glucose, LDL, and decrease in fasting insulin, HDL, HOMA-IR and  
477 HOMA-β was found. Similar bw and HDL results were found in a previous study  
478 conducted in our laboratory, but lower TC and TG levels was observed (Miranda et al.  
479 2018). These discrepancies may be attributed to variations in the experimental models  
480 and procedures.

481 Our data also showed that PM-Mothers presented an increase in insulin and TG  
482 levels in breast milk. The breast milk is considered crucial for the adequate development  
483 of the offspring. It includes several important substances, such as macronutrients  
484 (proteins, carbohydrates and fats), steroids, growth factors and hormones, such as leptin  
485 and insulin (Franco et al. 2012, Miranda et al. 2017, Ribeiro et al. 2017). Similar results  
486 was observed in breast milk from pregnant rabbits who were exposed to diluted and  
487 filtered diesel engine (DE) exhaust fumes from the 3<sup>rd</sup> to the 27<sup>th</sup> days post-conception  
488 (total exposure of 20 days out of a 31-day pregnancy) (Hue-Beauvais et al. 2019). Despite  
489 had not being possible to measure the difference in milk intake among the litters, we  
490 suggest that changes in the breast milk, from PM-Mothers, such as high levels of fat and  
491 insulin in its composition lead to exacerbation of the complications on SL-PM at PND 21  
492 and later life.

493 Epidemiological and experimental studies have consistently associating PM  
494 exposure in perinatal life to metabolic diseases such as T2DM and obesity in adult life  
495 (Haberzettl et al. 2016, Rao et al. 2015), since the PM maternal exposure can promote

496 low birth weight, increased adiposity, insulin resistance, and impairs glucose tolerance in  
497 adult offspring (Chen et al. 2017b). In addition, small litters are able to promote  
498 overnutrition by reduction in milk competition and increasing caloric intake (Ribeiro et  
499 al. 2017), as result, the SL animals present increased levels of circulating leptin and  
500 insulin resistance (Bei et al. 2015, Hou et al. 2011), companied with overweight or obesity  
501 phenotype throughout life (Wu et al. 2016).

502 In the present study the animals did not present difference at the birth weight.  
503 Similarly results also were found in mice from mothers treated with diesel exhaust PM<sub>2.5</sub>  
504 (DEP), from 7-weeks pre-conception until the weaning (Chen et al. 2017a). Nevertheless,  
505 at PND21 the SL PM showed an increase of fasting glucose and higher bw and fat  
506 deposition, as retroperitoneal and mesenteric fat pads (Table 3). Increased body weight,  
507 impaired whole-body glucose tolerance and decreased expression of insulin signaling  
508 genes in adipose tissue has also been found in mice exposure to PM<sub><2.5</sub> initiated at  
509 conception until 5 weeks of age, (Woodward et al. 2019).

510 The maternal treatment of PM<sub><10</sub> during pregnancy and lactation was not able to  
511 exacerbate insulin resistance rat offspring at PND 21, since less levels of fasting insulin  
512 as well decrease of HOMA-IR and HOMA-β index was observed. Taken together these  
513 data suggest that PM combined to SL model may lead to impaired insulin secretion in the  
514 β-cell, resulting in an increase of fasting glucose observed, in this sense, we could defend  
515 there was worsened the disease phenotype previously established.

516 Unlike PND 21, at PND 90 no statistical difference was found on the bw, body  
517 weight evolution (bwe), as well as in the fat pad stores, fasting glucose and fasting insulin,  
518 between SL-PM and SL-OIL (Table 3). On the other hand, at ivGTT, the SL-PM offspring  
519 showed a significantly increase of glucose plasma levels in the AUC, suggesting a glucose  
520 intolerance, same it had not observed hyperinsulinemia during the test. Curiously, an  
521 increment of insulin level was detecting in the first 5 minutes of the test compared to the  
522 SL-OIL offspring. Consistent with our findings, a pediatric cohort study, childhood  
523 elevated exposure to nitrogen dioxide (NO<sub>2</sub>) and PM<sub>2.5</sub> between the ages of 10 and 18,  
524 presented significant effects on insulin homeostasis, insulin sensitivity, and β-cell  
525 function. Although, they defend that PM exposure initially lead to an increase in insulin  
526 secretion, including higher fasting insulin levels and higher acute insulin response to  
527 glucose. The long-term PM exposure have been associated to β-cell fatigue and

528 consequently decrease in insulin secretion (Alderete et al. 2017). Thus, we assumed here  
529 that PM exposure was able to programs offspring, lead to a worsening in their disease  
530 phenotype as early as later in life.

531 In the current study, after a second challenge with HFD, from PND 90 to PND  
532 120, we did not observed difference at plasmas levels of glucose and insulin on the ivGTT  
533 between the groups, however, ipITT data clearly indicated insulin resistance in PM group  
534 at PND 120 compared to the OIL offspring, corroborating with HOMA-IR and HOMA-  
535  $\beta$  index. Similarly results was reported by Liu and colleagues in male offspring of Swiss  
536 Webster, with overnutrition until PDN 21 and after feeding with HFD between PDN 21-  
537 150, showed an disruption in the ability to regulate the food intake, increasing the  
538 sensitivity to HFD and increased the fat pad stores and adiposity (Liu et al. 2013). HFD  
539 is known by inducing increase bw, fat pad stores, fasting glucose, insulin resistance and  
540 total cholesterol, generally independent of animal age (Bock et al. 2016, Ibanez et al.  
541 2017).

542 Although it was not observed difference in food intake from PND 90 to PND 120  
543 between groups, SL-PM presented increased the body weight gain as well the  
544 retroperitoneal and mesenteric fat pad stores, after HFD. Curiously, mice that fed HFD  
545 by 16 weeks, presented a decrease on hyperglycemia after to high-intensity exercise,  
546 however, when they were exposure to  $PM_{<2.5}$  that reduction was not observed (Kostrzycki  
547 et al. 2019), suggesting a combined effect of PM and HFD that lead to an increase of  
548 adiposity in SL-PM group.

549 Recent results in this field have been shown that chronic PM exposure combined  
550 with HFD cause a synergistic effect impairing oxidative homeostasis, changes of lipid  
551 accumulation, oxidative stress, inflammation and hepatic steatosis in mice (Ding et al.  
552 2019). In our study the assessment of the oxidative status of the liver from offspring at  
553 PND 120, demonstrated a significative reduction of the levels of SOD and GST. And  
554 lower levels of GSH were found. These enzymes are both oxidative stress markers and  
555 play an important role to protect against the cellular damage, while SOD is able to  
556 eliminate superoxide radicals converting, GST may acting detoxifying xenobiotic  
557 compounds and lipidic hydroperoxides, in addition, decrease of GSH represents lower  
558 cell capacity to eliminate reactive species and peroxides (Borges et al. 2018). This way,

559 a decrease on SOD activity may be suggest a lower capacity to eliminate reactive oxygen  
560 species due to damage to cell machinery.

561 In previous studies with squamous cell carcinoma of the nasal septum human, the  
562 PM<sub>2.5</sub> was able to increase inflammatory response and decrease the activities of SOD,  
563 catalase and glutathione peroxidase (Hong et al. 2016). In another finding, PM<sub>2.5</sub>  
564 exposure, for 48 hours, in HBE and BEAS-2B cells was able to increase reactive oxygen  
565 species (ROS) levels and IL-6 secretion in this cell culture. Notably, IL-6 is known as  
566 important mediator and a strong indicator of the inflammatory response (Yuan et al.  
567 2019). In addition, six-week-old C57BL/6 male mice that were exposed to concentrated  
568 ambient PM<sub>2.5</sub> by 60 days, presented an increase in endoplasmic reticulum stress and  
569 inflammatory status in the lung and liver (Laing et al. 2010). However, when the  
570 C57BL/6 male mice were fed HFD and after exposure to concentrated ambient PM<sub>2.5</sub>, it  
571 was observed markedly worsened insulin and glucose homeostasis, besides increase in  
572 inflammation (Sun et al. 2009).

573 Indeed, the liver is considered the main organ and a critical place of lipid  
574 metabolism (Xu et al. 2019), so, although our data did not presented difference in lipid  
575 peroxidation and Myeloperoxidase, we observed that SL-PM group presented alteration  
576 of the lipid profile at all ages evaluated. Curiously, mice that were exposure to different  
577 concentrations of PM<sub>2.5</sub> for 6 months, showed that genes related to lipid accumulation  
578 who was resulting with increases in liver TG, TC and VLDL levels, followed by IR,  
579 glucose intolerance and abnormal blood pressure. In this sense is clearly there is a strong  
580 association among an oxidative stress, inflammation and insulin resistance (Giacco  
581 & Brownlee 2010, Xu et al. 2019) and as we could observed, several studies shown that  
582 PM exposure might lead to oxidative stress, inflammation and consequently development  
583 of the insulin resistance (Park 2017, Thiering et al. 2013).

584 In fact, the animals evaluated in this work were early overfeed and submitted to  
585 the HFD in adult life, being these consistent models for studies of metabolic dysfunctions.  
586 Thus, our major aim in the current study was to test the hypothesis that maternal  
587 exposition to PM<sub><10</sub> during gestation and lactation could exacerbate dysfunctions already  
588 found on offspring metabolism, as short- and long-term consequence. Although, this  
589 study has some limitations about a better explanation upon the mechanism behind  
590 malprogramming consequences from PM<sub><10</sub>, further experiments with this model are

591 essential to verify the mechanisms by which PM programming exacerbate the  
592 dysfunctions on insulin and lipid metabolism in mothers and offspring and oxidative  
593 stress in the offspring at adulthood. Nevertheless, our results present very important  
594 evidences to hypothesis confirmation that environmental pollutants exposure during  
595 perinatal period combined with early overnutrition and HFD at adulthood worsened  
596 metabolic dysfunction in the descendants.

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615 **5. Conclusion**

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617 In summary, we demonstrate that maternal exposure to PM<sub><10</sub>, collected from a  
618 high-density-traffic urban area, during gestation and lactation, was able to exacerbate  
619 dysfunctions on metabolism in obese animal model offspring, increasing glucose  
620 intolerance and dyslipidemia later in life. Furthermore, after a nutritional challenge with  
621 HFD at adulthood, metabolic dysfunctions become more evident, demonstrating that PM  
622 exposure in early life worsens when combined with nutritional insults, increasing  
623 oxidative stress, obesity and leading to dyslipidemia and insulin resistance at adulthood.  
624 While previous animal studies have yielded similar results, our findings are the first to  
625 show the effect of programming of PM<sub><10</sub> on recognized obesity models, such as early  
626 overnutrition combined with HFD at adulthood.

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641 **Declaration of interest**

642 The authors declare no conflict of interest associated with this manuscript.

643

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647

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649

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889 **Table 1. Effects of PM on biometric and biochemical parameters of mothers at weaning**

Parameters	OIL-Mothers	PM-Mothers
AUC bw gestation	5985 ± 209.9	5787 ± 177.4
AUC bw lactation	6184 ± 194.4	6121 ± 151.3
Retroperitoneal fat pad (g/100 g bw)	1.41 ± 0.12	1.01 ± 0.14
Ovarian fat pad (g/100 g bw)	0.84 ± 0.15	0.70 ± 0.14
Uterine fat pad (g/100 g bw)	1.39 ± 0.16	1.02 ± 0.15
Mesenteric fat pad (g/100 g bw)	0.72 ± 0.07	0.69 ± 0.08
Fasting glucose (mg/dL)	97.4 ± 3.22	111.6 ± 1.12**
Fasting insulin (ng/dL)	0.122 ± 0.01	0.061 ± 0.01**
HOMA-IR	0.65 ± 0.08	0.39 ± 0.06*
HOMA-β	29.96 ± 2.81	9.73 ± 2.16****
Total cholesterol (mg/dL)	93.49 ± 4.37	82.91 ± 4.2
HDL cholesterol (mg/dL)	51.7 ± 3.01	34.7 ± 1.03****
VLDL cholesterol (mg/dL)	10.42 ± 0.23	10.7 ± 0.81
LDL cholesterol (mg/dL)	32.52 ± 1.36	39.89 ± 2.18*
Triglycerides (mg/dL)	53.9 ± 2.06	57.3 ± 3.79

890 Data are expressed as the mean ± SEM; \*p < 0.05, \*\*p < 0.01, \*\*\*p < 0.001 and \*\*\*\*p < 0.0001  
 891 by Student's *t* test (n = 7 rats/group).

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903 **Table 2. Effects of PM on milk biochemical parameters at weaning.**

Parameters	OIL-Mothers	PM-Mothers
Milk glucose (mg/dL)	190.1 ± 11.81	200.1 ± 16.58
Milk insulin (ng/mL)	0.768 ± 0.09	2.054 ± 0.114****
Milk triglycerides (mg/dL)	931.7 ± 51.25	1138.0 ± 70.77*
Milk total cholesterol (mg/dL)	85.3 ± 8.11	73.18 ± 6.97

904 Data are expressed as the mean ± SEM; \*p < 0.05, \*\*\*\*p < 0.0001 by Student's t test (n = 7  
905 rats/group).

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927 **Table 3. Effects of maternal PM treatment on the biometric and biochemical parameters in**  
 928 **offspring at PND 21 and PND 90**

Parameters	SL-OIL	SL-PM
<b>PND 21</b>		
Fasting glucose (mg/dL)	103.2 ± 3.89	114.8 ± 3.46*
Fasting insulin (ng/dL)	0.1978 ± 0.03	0.0873 ± 0.03**
HOMA-IR	1.15 ± 0.09	0.63 ± 0.14**
HOMA-β	34.44 ± 2.69	17.35 ± 3.58***
Total cholesterol (mg/dL)	69.92 ± 3.07	68.62 ± 3.75
HDL cholesterol (mg/dL)	25.15 ± 0.89	21.5 ± 0.92**
VLDL cholesterol (mg/dL)	7.8 ± 0.31	11.23 ± 0.87***
LDL cholesterol (mg/dL)	40.22 ± 1.79	41.43 ± 2.83
Triglycerides (mg/dL)	39.0 ± 1.54	56.17 ± 4.36***
<b>PND 90</b>		
Fasting glucose (mg/dL)	78.8 ± 1.7	84.1 ± 2.0
Fasting insulin (ng/dL)	0.329 ± 0.03	0.327 ± 0.02
HOMA-IR	1.406 ± 0.09	1.427 ± 0.05
HOMA-β	216.7 ± 2.69	137.2 ± 3.58*
Body Weight (g)	371.8 ± 5.76	389.9 ± 11.51
Retroperitoneal fat pad (g/100 g bw)	1.70 ± 0.06	1.54 ± 0.08
Mesenteric fat pad (g/100 g bw)	0.829 ± 0.04	0.723 ± 0.05
Total cholesterol (mg/dL)	61.06 ± 5.76	91.06 ± 5.65**
Triglycerides (mg/dL)	53.58 ± 2.22	83 ± 13.37*

929 Data are expressed as mean ± SEM; \*p < 0.05, \*\*p < 0.01 and \*\*\*p < 0.001 by Student's t test  
 930 (n = 6/group from 6 different litters).

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936 **Table 4. Effects of maternal PM treatment on the biometric and biochemical parameters in**  
 937 **offspring at PND 120 under the HFD treatment**

Parameters	SL-OIL	SL-PM
Body Weight (g)	484.5 ± 9.80	466.5 ± 7.48
Retroperitoneal fat pad (g/100 g bw)	2.92 ± 0.05	3.28 ± 0.14*
Mesenteric fat pad (g/100 g bw)	1.19 ± 0.08	1.49 ± 0.08*
Fasting glucose (mg/dL)	75.9 ± 6.42	75.2 ± 2.26
Fasting insulin (ng/dL)	0.438 ± 0.07	0.658 ± 0.08
Total cholesterol (mg/dL)	78.84 ± 2.17	99.06 ± 4.57***
Triglycerides (mg/dL)	61.0 ± 3.8	58.3 ± 4.1
HOMA-IR	1.791 ± 0.35	2.985 ± 0.32*
HOMA-β	210.3 ± 30.9	501.8 ± 110.8*

938 Data are expressed as the mean ± SEM; \*p < 0.05, \*\*p < 0.01 and \*\*\*p < 0.001 by Student's t  
 939 test (n = 6/group from 6 different litters).

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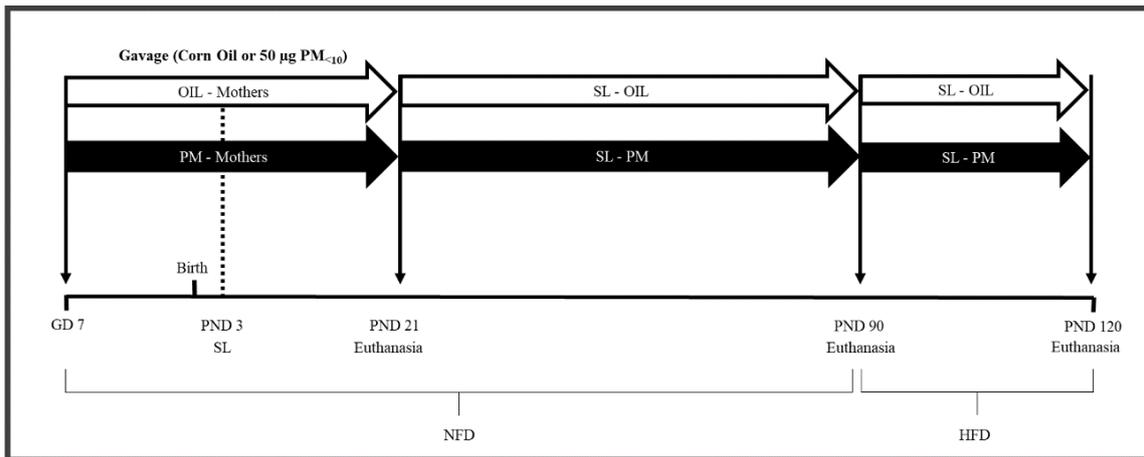
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955 **Figure 1. Experimental design:** OIL - Mothers, mothers who received corn oil; PM-Mother,  
 956 mothers who received PM<sub>10</sub> solution; SL - OIL, offspring from OIL - Mothers; SL - PM,  
 957 offspring from PM - Mothers; PND, post-natal day; NFD, normal fat diet; HFD, high fat diet.

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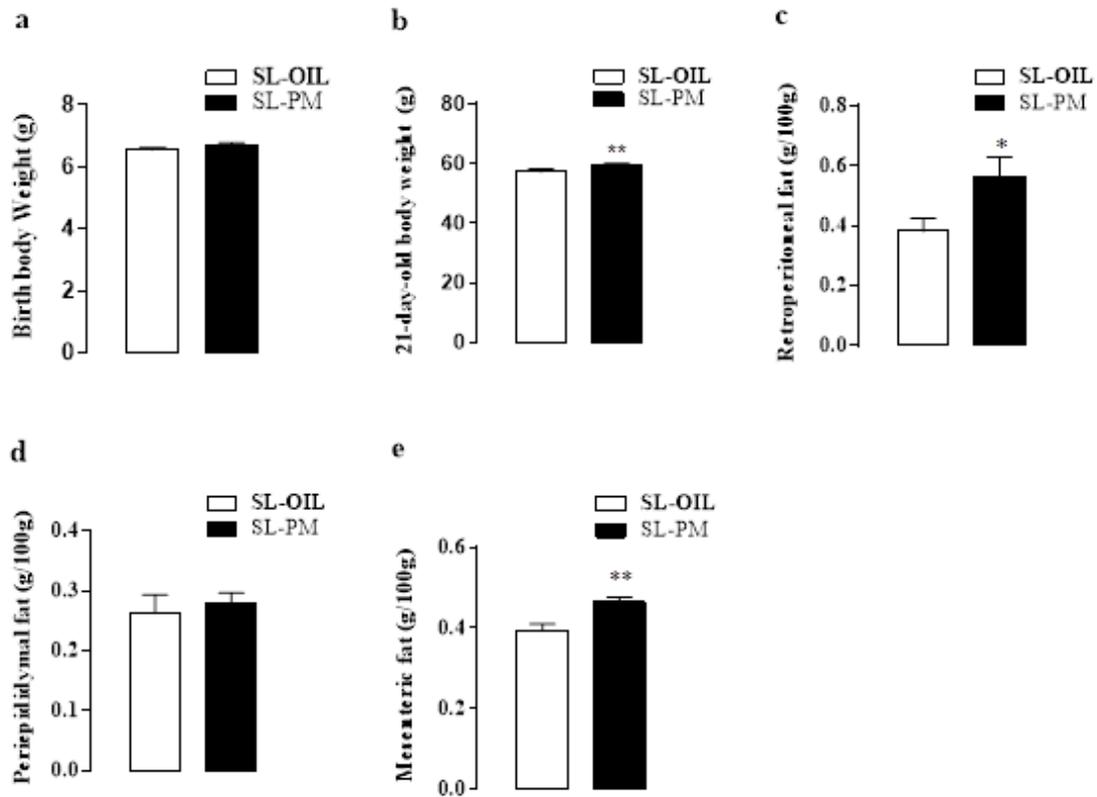
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974 **Figure 2. Effect of PM<sub><10</sub> during gestation and lactation on birth weight, body weight and**  
 975 **tissue fat accumulation in the offspring at PND 21.** Birth weight in OIL and PM offspring (a)  
 976 (n = 21/group from 7 litters), body weight at PND 21 (b) (n=21/group from 7 litters),  
 977 retroperitoneal (c) (n = 6-9/group from 6 different litters), periepididymal (d) (n = 6-9//group from  
 978 6 different litters) and mesenteric fat pads (e) (n = 6-9//group from 6 different litters). Data are  
 979 expressed as the mean ± SEM; \*p < 0.05, \*\*p < 0.01 by Student's t test.

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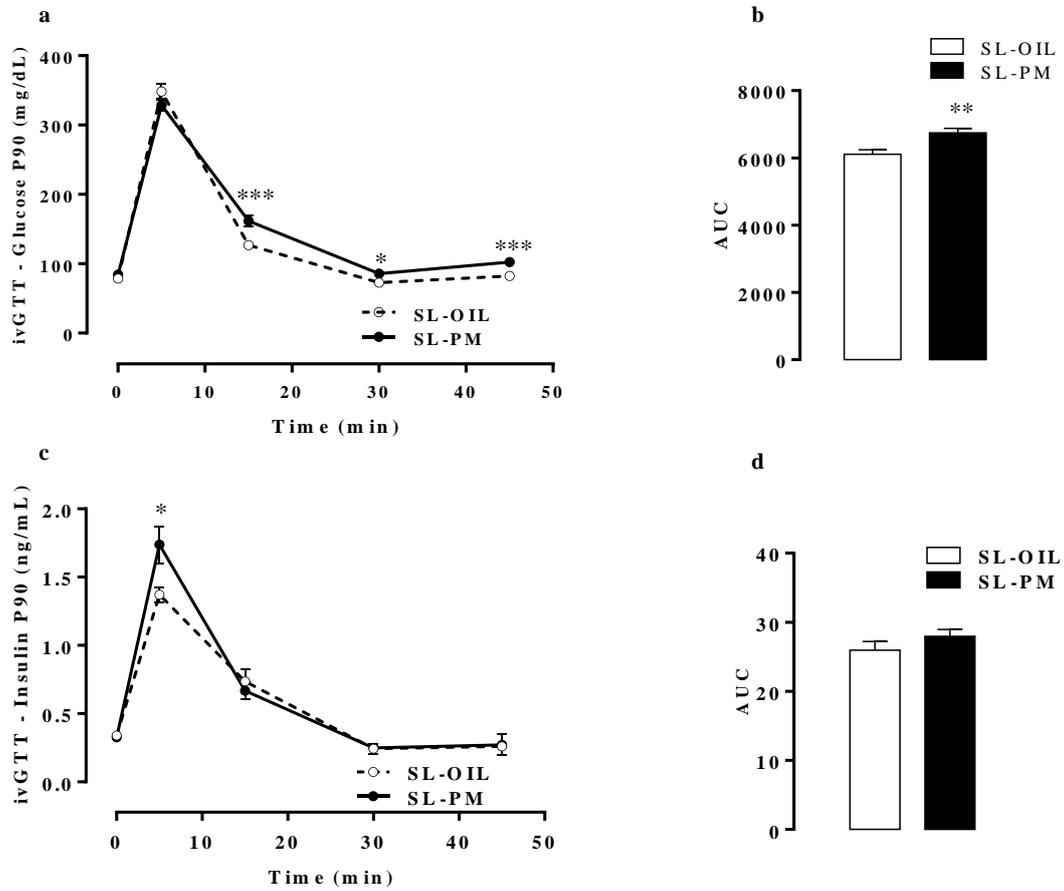
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993 **Figure 3. Effect of  $PM_{10}$  on plasma glucose and insulin levels during ivGTT in PND 90**  
994 **offspring.** Glucose (a) and insulin (b) levels in ivGTT. Data are expressed as mean  $\pm$  SEM; \* $p$   
995  $< 0.05$ , \*\* $p < 0.01$ , \*\*\* $p < 0.001$  by Student's  $t$  test ( $n = 6$ /group from 6 different litters).

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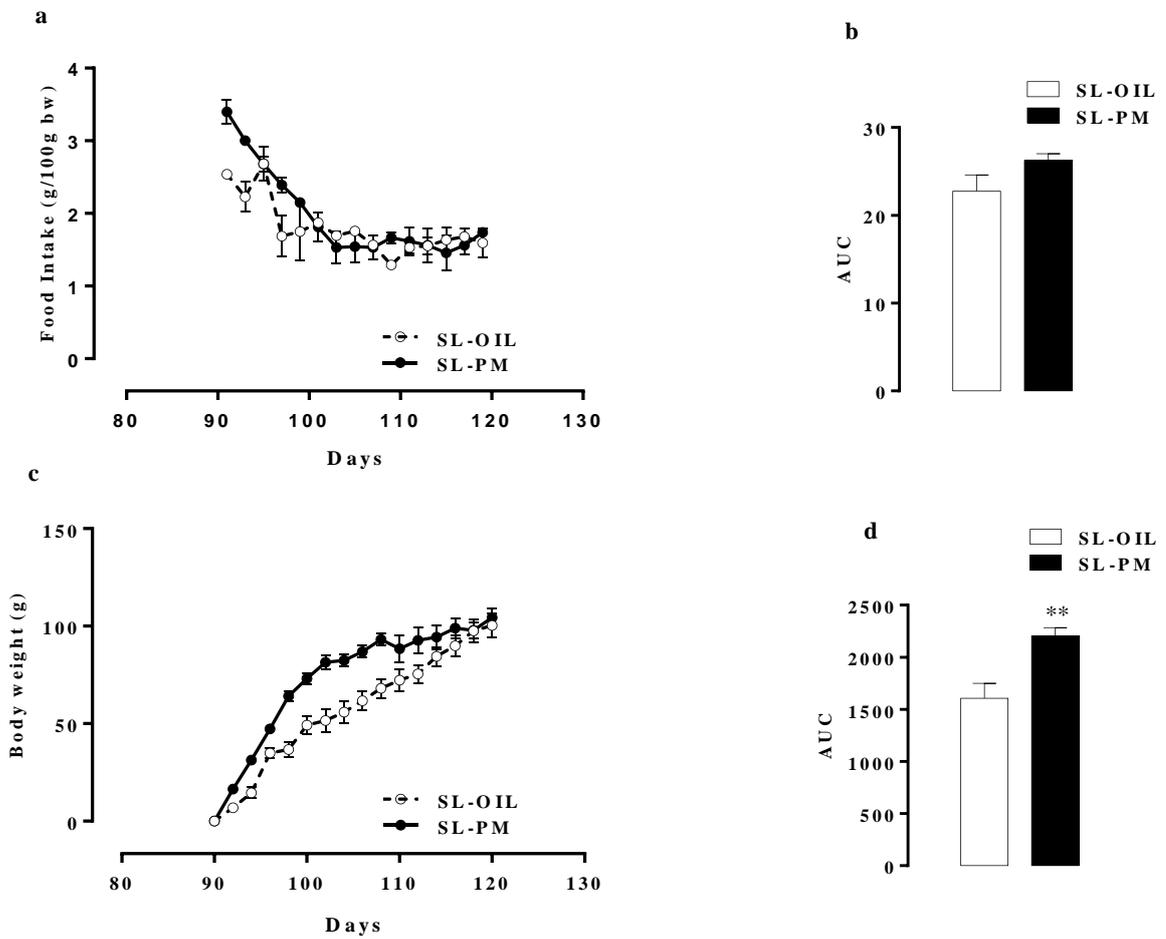
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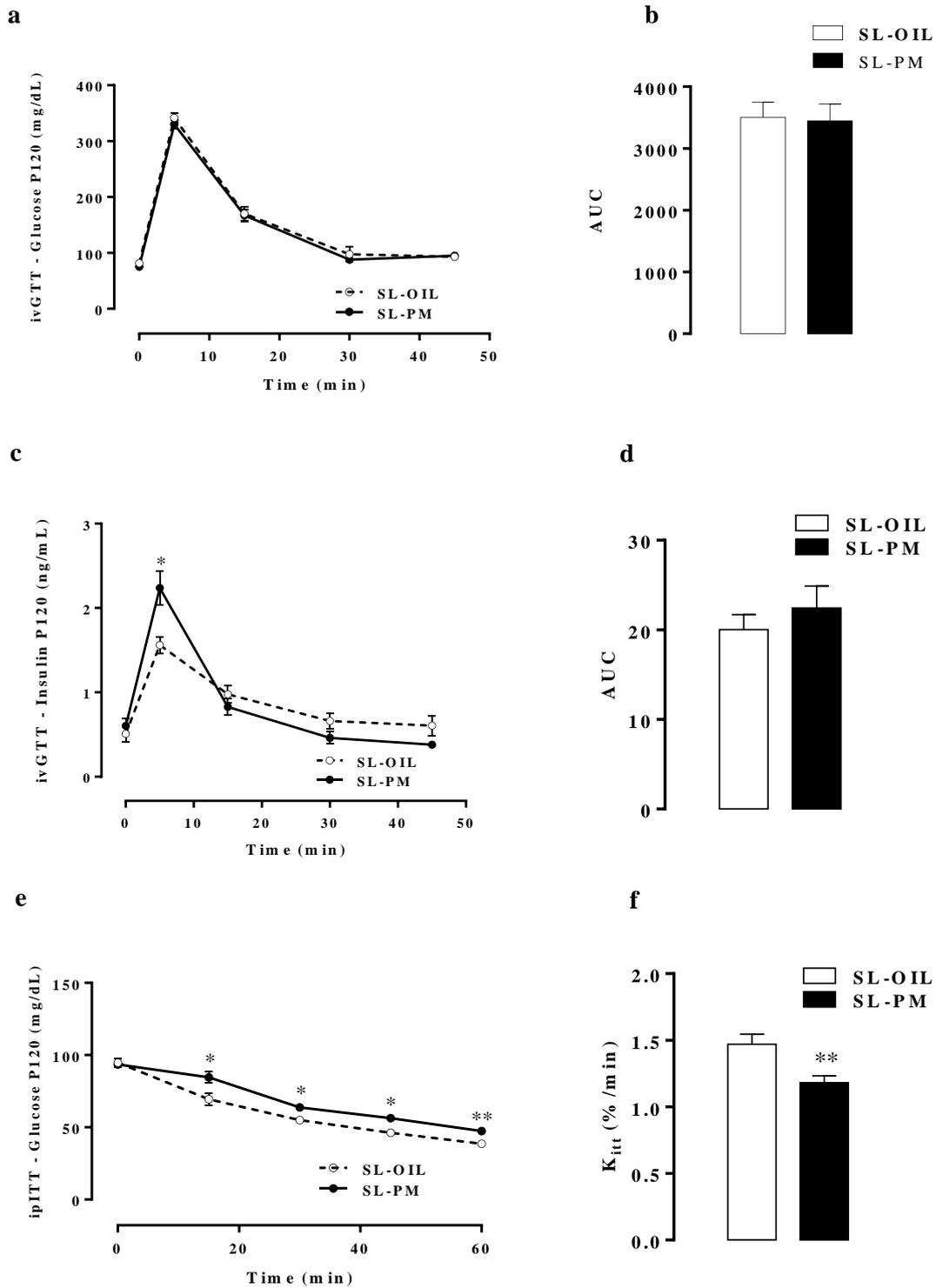
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1007 **Figure 4. Effect of PM<sub>10</sub> during gestation and lactation on body weight evolution and food**  
1008 **intake of the offspring from PND 90 until PND 120.** Body weight (a), AUC of body weight  
1009 (b), Food intake (c) and AUC of food intake (d). Data are expressed as mean ± SEM; p < 0.05 by  
1010 Student's *t* test (n = 6/group from 6 different litters).

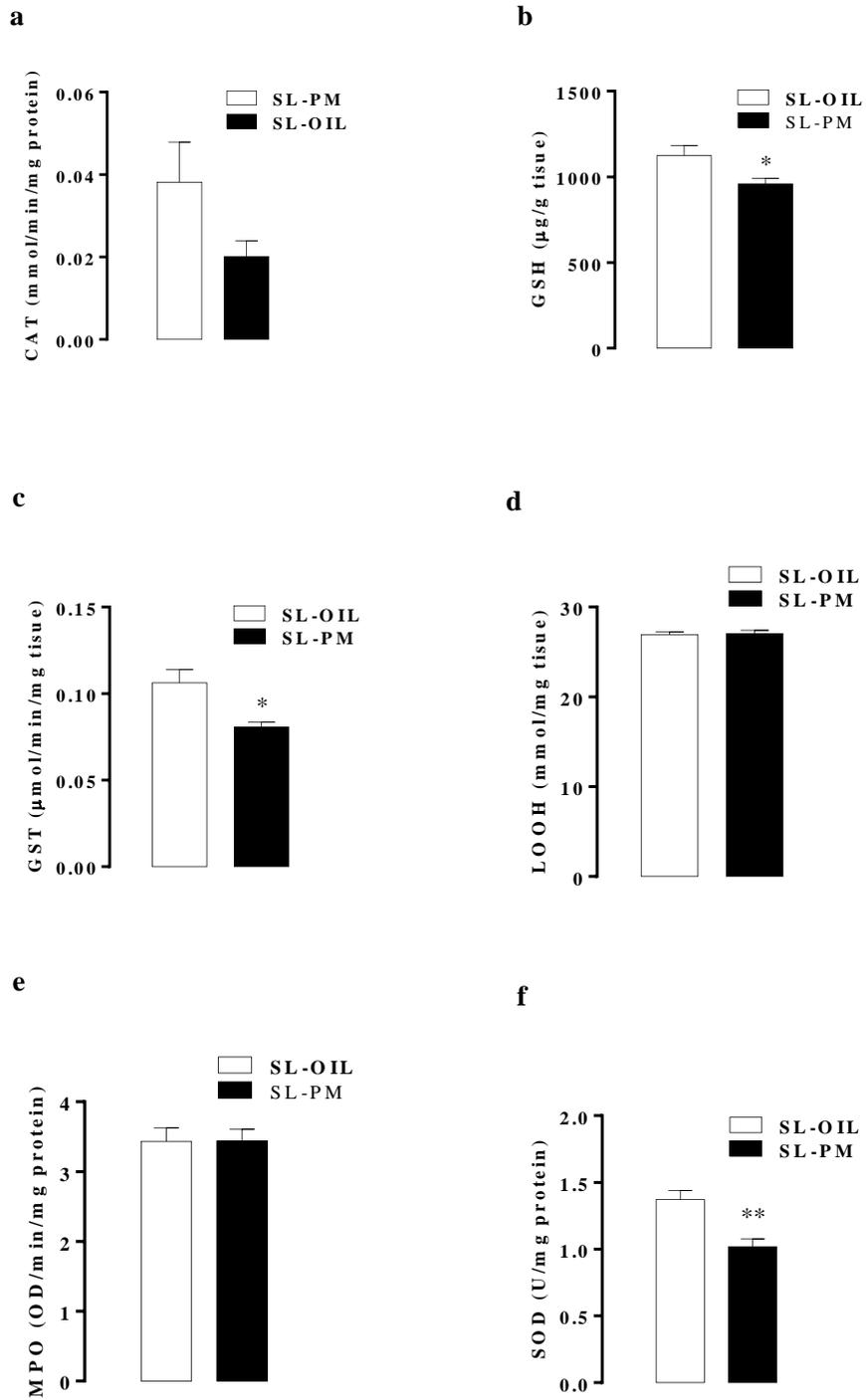
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1013 **Figure 5. Effects of PM<sub>10</sub> on glucose and insulin plasma during ivGTT and glucose plasma**  
 1014 **in ipITT in PND 120 offspring.** Glucose (a) and insulin (b) levels in ivGTT .Glucose level (c)  
 1015 in ipITT. Data are expressed as mean ± SEM; \*p < 0.05, \*\*p < 0.005 by Student's *t* test (n =  
 1016 6/group from 6 different litters).

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1020 **Figure 6. Effect of PM<sub>10</sub> on oxidative stress and myeloperoxidase in PND 120 offspring.**

1021 Catalase (a), GSH: glutathione (b), GST: glutathione transferase (c) LOOH: lipid hydroperoxide

1022 (d), MPO: myeloperoxidase (e) and SOD: Superoxide Dismutase (f). Data are expressed as mean

1023 ± SEM; \*p < 0.05, \*\*p < 0.005 by Student's *t* test (n = 6/group from 6 different litters).

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1025 **High fat diet during adolescence in male rats programs cardiometabolic**  
1026 **dysfunctions in adulthood**

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1058 **Background/aims:** There is a negative relationship between exposure to high fat diet  
1059 (HFD) in early life and changes in blood pressure in adulthood. It has also been suggested  
1060 that adolescence is a susceptible phase for programming to metabolic syndrome (MetS).  
1061 Thus, our hypothesis argues that HFD during adolescence can lead to cardiometabolic  
1062 dysfunctions at adulthood. **Methods:** Adolescent Wistar rats (30 to 60 day-old) were  
1063 exposed to a HFD (35% of fat). Control animals had access to normal commercial chow  
1064 (NFD, 4.5% of fat). Glycemia, lip profile, food intake, body weight, fat pad stores, blood  
1065 pressure, heart rate and pulse pressure were verified in 120-day-old rats. Student t-test  
1066 was used to compare data between groups. **Results:** At basal test, HFD animals showed  
1067 increased systolic blood pressure (SBP) and mean blood pressure (MBP) compared with  
1068 control animals (SBP:  $125.4 \pm 1.8$  mmHg vs.  $117 \pm 1.9$  mmHg, respectively,  $p < 0.05$ ;  
1069 MBP:  $95.78 \pm 1.8$  vs.  $88.85 \pm 1.6$  mmHg, respectively,  $p < 0.05$ ). Diastolic blood pressure  
1070 (DBP) was similar between groups, as well as pulse pressure and heart rate. Furthermore,  
1071 was observed an increase in body weight gain, as well fat pad stores, hypertriglyceridemia  
1072 and glycemia in the intravenous glucose tolerance test (ivGTT). Blood pressure decrease  
1073 in response to hexamethonium injection (30mg/kg of body weight) was greater in HFD  
1074 animals compared with control animals ( $\Delta$ SBP  $-43.5 \pm 3.1$  vs.  $-33.8 \pm 3.1$  mmHg;  $\Delta$ MBP  
1075  $-37.9 \pm 3.3$  vs.  $-27.8 \pm 1.9$  mmHg;  $\Delta$ DBP  $-33.2 \pm 3.5$  vs.  $-23.7 \pm 1.1$  mmHg respectively,  
1076  $p < 0.05$ ). No difference between groups was observed in pulse pressure (HFD:  $-10.2 \pm$   
1077  $2.1$  and NFD  $-9.4 \pm 2.2$  mmHg,  $p=0.4$ ) and in heart rate response (HFD:  $-5.9 \pm 20.2$  and  
1078 NFD  $-3.3 \pm 6.9$  bpm,  $p=0.2$ ) in response to hexamethonium. **Conclusions:** HFD during  
1079 adolescence was able to program rats to development MetS, leading to higher levels of  
1080 adiposity, hypertriglyceridemia, SBP and MBP later in life. In addition, the exacerbated  
1081 blood pressure decrease in response to hexamethonium, points to higher sympathetic  
1082 activity in HFD rats than in control ones, taken together these data suggest that pubertal  
1083 period is an important programming window.

1084  
1085 **Keywords:** Adolescence, metabolic disease, high-fat diet.

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1096 **1. Introduction**

1097 Metabolic syndrome (MetS) has been main cause of morbidity and mortality. In  
1098 addition, presenting a close association with obesity, type 2 diabetes mellitus (T2DM)  
1099 and cardiovascular disease (CVD), the prevalence of MetS and obesity together have  
1100 reaching an epidemic status in worldwide [1, 2]. It has been estimated that number of  
1101 overweight or obese among adult people is about 40% [3]. In this context, the high  
1102 prevalence of MetS may be due the changes in the lifestyle in the last decades,  
1103 including lower levels of physical activity, increased in adiposity and socioeconomic  
1104 status [4]. Furthermore, other factors as smoking, difference in genetic inheritance,  
1105 cases of T2DM on family and education seems to have a strong association with MetS  
1106 and its other diseases associated [5]. Although there is no a single definition to MetS,  
1107 it may be defined as a cluster of three or more of the following risk factors together:  
1108 obesity, glucose intolerance, hypertension and dyslipidemia, specifically  
1109 hypertriglyceridemia [1, 6].

1110 Among the several risk factors associated to MetS and obesity, the changes in diet  
1111 composition seems to have a key role to development of obesity and consequent  
1112 MetS. In this sense, diet with high fat content (HFD) are often related to increase of  
1113 body weight (bw) by increase in fat pad stores, both in humans and experimental [7].  
1114 Furthermore, this diet lead to high level of fasting glucose, total cholesterol and  
1115 insulin resistance (IR), generally independent of exposure age [8, 9].

1116 Several studies have shown that gestation and lactation are critical periods of body  
1117 development, thus, it has been suggested that exposure to environmental disruptors,  
1118 during early life, could lead to programming of the offspring to cardiometabolic  
1119 disease later in life [10-13]. This exposure may lead to noncommunicable disease,  
1120 such as obesity, coronary heart disease, T2DM, and some types of cancer at adulthood  
1121 [10, 11]. Corroborating this findings, previous study of our group have show that  
1122 offspring from mothers exposure to HFD during gestation and lactation presented  
1123 increase in bw and adiposity at weaning, in addition these pups showed  
1124 hyperglycemia, hyperleptinemia and elevated blood pressure at adulthood [14, 15].

1125 Recent studies have shown that peri-pubertal period also is critical period for body  
1126 development as well of the sexual maturation and development of central nervous  
1127 system (CNS) [9, 11, 16]. In addition, HFD was able to induce changes in both

1128 reproductive system and metabolism, and consequently led to metabolic dysfunctions  
1129 at adulthood [9]. Nevertheless little is known about the effect of HFD during  
1130 adolescence and its role on cardiometabolic syndrome later in life. Thus, we  
1131 hypothesized that consumption of HFD during peri-pubertal period is able to  
1132 programing changes and induce to cardiometabolic syndrome at adulthood.

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1152 **2. Materials and Methods**

1153 2.1. Experimental model and diet

1154 Twenty-five day-old male Wistar rats were supplied by the Central Animal House  
1155 at the State University of Maringá, Paraná and kept in the Animal House of Department  
1156 of Biotecnology, Genetics and Cellular Biology in light controlled conditions with a 12-  
1157 h light-dark cycle (07:00 a.m. to 07:00 p.m.) and temperature of  $22.0 \pm 2$  °C. After five  
1158 days of adaptation, the animals were randomly distributed in five per cage. At 30-days-  
1159 old a group of animals were fed with high fat diet (HFD; hypercaloric diet with 35% lard/  
1160 5.817 kcal/g) [17] for thirty days (HFD group). After that, the rats were switched to  
1161 standard diet containing an adequate amount of protein (20.5% protein, Nuvital®,  
1162 Curitiba/PR, 3.801 kcal/g) (NFD group). Controls animals were fed with standard diet  
1163 during all protocol. Water and food were provided *ad libitum* in both groups.  
1164 Experimental assays were performed with 15 animals per group and the protocol was  
1165 approved by the Ethics Committee of the State University of Maringá (protocol number  
1166 1527130815).

1167 2.2. Food intake, body weight, evolution and fat tissue accumulation (biometric and  
1168 biochemical parameters)

1169 Between 30 and 120 days-old, the body weight and food intake were assessed,  
1170 once and three times a week, respectively. The average of food intake was calculated per  
1171 rat per day and expressed relative to 100g of body weight. At 120-days of age, a group of  
1172 animals were euthanized by decapitation method and fat pad stores was removed and  
1173 weighed, to measure fat tissue accumulation. Plasma was used for quantify, fasting  
1174 glucose, total cholesterol and triglycerides by enzymatic method using a specific  
1175 colorimetric commercial kit (Gold Analisa®, Belo Horizonte, Minas Gerais, Brazil).

1176 2.3. Intravenous glucose tolerance test (ivGTT)

1177 At PND 120 the offspring (n=6 for each group and age) underwent surgery  
1178 consisting of silicone cannula implantation into the right jugular vein as previously described  
1179 (de Oliveira et al. 2011). After 12-h fasting, a glucose load (1 g/kg bw) was injected into the  
1180 cannula. Blood samples (400 µl) were collected immediately prior to glucose injection (0  
1181 min) and then at 5, 15, 30 and 45 min, and the obtained plasma samples were stored at -20

1182 °C for further analysis. To maintain blood volume, a corresponding volume of saline (0.9%)  
1183 was infused through the cannula.

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### 1185 2.3. Surgery for arterial catheter implantation

1186 At 120 days of life, another group of the rats were submitted to intramuscular  
1187 anesthesia (Ketamine-xylazine; 3 and 0.6 mg/100g of body weight, respectively) and a  
1188 P10 catheter (P10 cannula-Micro-Renathane linked to a P50 cannula-ClearTygon) that  
1189 was filled with 0.1 ml of heparinized saline (500 units/mL) was installed into the femoral  
1190 artery and advanced (4 cm) until the tip entered the abdominal aorta. During surgery a  
1191 Doxycycline dose (2 mg/kg of body weight, intra-arterial) was administered and the next  
1192 two days after surgery analgesic metamizole was provided (30mg/kg). After surgery,  
1193 animals were housed in individual cages. To eliminate any clot, 0.1 ml of heparinized  
1194 saline (500 units/mL) was injected through the cannula [18] and blood pressure  
1195 recordings were performed four days later [19].

### 1196 2.4. Protocol and experimental procedures for blood pressure assessment

1197 After 1 hour of adaptation the experiments were performed in 3 animals from NFD  
1198 and HFD groups running in parallel. It was wait thirty minutes in adaptation to allow  
1199 blood pressure stabilization and a baseline recording during 30 minutes, when animals  
1200 were relaxed and quiet or sleeping. All protocol were performed during the inactive  
1201 period of animals, between 1 and 4 p.m. [20]. Following, a dose of Hexamethonium (30  
1202 mg/kg) was injected intraperitoneally and blood pressure recording for subsequent 30  
1203 minutes.

### 1204 2.5. Measurement of blood pressure and heart rate

1205 The arterial cannula was connected to a fluid-filled blood pressure transducer  
1206 (MLT0699, AD Instruments, Dunedin, New Zealand), which was linked to a signal  
1207 amplifier (Insight, Ribeirão Preto/SP Brazil). Thus, continuous recordings of arterial  
1208 pressure were sampled at 1000Hz using an analog-to-digital converter board (CODAS,  
1209 1-kHz sampling frequency, Dataq Instruments, Inc, Akron, OH) with freely moving rats  
1210 in their home cage. To measure systolic, diastolic and calculated mean arterial pressure  
1211 (SBP, DBP and MBP, respectively) and pulse interval (PI; a surrogate for the R-R  
1212 interval) values. Analyses were made on a beat-to-beat basis [21] over 20 min and during  
1213 Hexamethonium test.

1214 2.6. Statistical analyses

1215 Statistical analyses and graph design were performed using GraphPad Prism  
1216 version 6.01 for Windows (GraphPad Software, La Jolla, CA, USA), being the data  
1217 expressed as mean  $\pm$  SEM. Students t-test was used and p value  $< 0.05$  was considered as  
1218 statistical significant differences between groups.

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### 1238 3. Results

#### 1239 3.1 Food intake, body weight evolution and fat tissue accumulation

1240 HFD adolescent rats showed a decreased food intake during the diet period  
1241 treatment compared to control animals, however no difference was observed after this  
1242 period between groups (Fig. 1A,  $p < 0.0001$ ). In addition, the content of calories intake  
1243 was higher in the HFD group only during the diet period and no difference was verified  
1244 in calories intake after that (Fig. 1B,  $p < 0.005$ ). Furthermore, an increase in the weight  
1245 gain was observed during and after HFD treatment (23.9%,  $p < 0.01$ ; 14.7%,  $p < 0.01$ ,  
1246 respectively; Fig. 1C). Besides, an increase of bw, the retroperitoneal, periepididymal and  
1247 mesenteric fat pad stores was observed, compared to controls (14.7%,  $p < 0.01$ ; 54.7%,  $p$   
1248  $< 0.01$ ; 36.8%,  $p < 0.01$  and 56.2%,  $p < 0.01$ , respectively; Fig. 2).

#### 1249 3.2 Lipid profile and glucose homeostasis

1250 Although the total cholesterol did not show significant difference between the  
1251 groups ( $p = 0.0602$ ), triglyceride levels were elevated in HFD animals (59.5%,  $p < 0.001$ ;  
1252 Fig. 3). As indicated in the Figure 4, an increase of plasma glucose levels was observed  
1253 in HFD animals at the intravenous glucose tolerance test (ivGTT) in the AUC compared  
1254 to controls (13.5%,  $p < 0.05$ ), this increase of glucose plasma levels was observed  
1255 mainly at the five minutes after glucose injection in HFD group (40.6%,  $p < 0.01$ ).

#### 1256 3.3 Cardiovascular measurements

1257 Cardiovascular analyses during resting period did not show difference in pulse  
1258 interval, heart rate and pulse pressure, animals exposed to HFD during adolescence  
1259 presented an increase in SBP and MBP in adulthood (9.3%,  $p < 0.05$  and 7.1%,  $p < 0.05$ ,  
1260 respectively). Representative signals of blood pressure recordings of controls and HFD  
1261 animals are shown in Fig 5.

#### 1262 3.4 Cardiovascular response to hexamethonium

1263 After hexamethonium exposure, an expressive decrease in the DBP, SBP and  
1264 MBP (35%,  $p < 0.05$ ; 19%,  $p < 0.01$  and 30.3%,  $p < 0.05$ , respectively) was observed in  
1265 HFD group compared to control group. However, differences on heart rate were not  
1266 statistically significant.

#### 1267 **4. Discussion**

1268 In this study HFD adolescent rats showed changes in calorie intake during the diet  
1269 period treatment, as well in the weight gain, lipid profile and cardiovascular parameters  
1270 at adulthood. Taken together, these findings suggest that chronic HFD exposure during  
1271 adolescence, have long term consequences in health and programs the development of  
1272 cardiometabolic syndrome later in life. Gestation, lactation and adolescence are  
1273 particularly crucial periods for brain structures development. Thus, several of studies  
1274 have shown that the exposure to environmental stressors in each one of these periods  
1275 could be responsible to development of metabolic diseases at adulthood, pointing to the  
1276 developmental origins of health and disease (DOHaD) concept [9-13].

1277 Male rats fed with HFD by thirty days, during adolescence, presented changes on  
1278 cardiometabolic parameters at 120 days of life signaling to increased risk of  
1279 cardiovascular disease and MetS. These findings agree with previous reports [9]. It has  
1280 been discussed that deleterious effect of insults during adolescence are both related to  
1281 behavior and metabolic changes at adulthood, pointing to the sensibility of central  
1282 nervous system (CNS) during adolescence due its maturation [16, 22]. In this sense, the  
1283 expressively increase of weight gain, mainly after the HFD period, may be attributed to  
1284 less energy expenditure observed in HFD animals, since the HFD exposure is related to  
1285 decrease of mobility [22, 23]. Indeed HFD exposure has been related to damage on  
1286 arcuate nucleus from hypothalamus [24], and, as it has already been widely known, that  
1287 food intake, weight gain and metabolic parameters are regulated by orexigenic (anabolic)  
1288 and anorexigenic (catabolic) neuropeptides produced by hypothalamic regions, that are  
1289 related and integrated with satiety signals like glucose, insulin and leptin [25]. However,  
1290 this study does not discuss that into metabolic programming context. On the other hand,  
1291 studies about maternal programming by HFD exposure have shown that mice offspring  
1292 presents increased changes in food intake as well as increased body weight at adulthood  
1293 [26, 27]. Furthermore, our group showed that HFD during perinatal life led to  
1294 hyperleptinemia and leptin resistance in the offspring at weaning, and these effects are  
1295 strongly associated with higher body weight and adiposity at adulthood [14].

1296 Notably, a previous study of our group discussed that HFD exposure at  
1297 peripubertal phase led to more drastic consequences than HFD exposure at adult life.  
1298 Thus, it has been suggested that adolescence as well gestation and lactation periods are

1299 susceptible programming window, where some regions that are responsible by food  
1300 intake and body weight regulation, e. g. hypothalamus, might be seriously affected and  
1301 consequently promote changes in metabolism [17]. In this context, organization problems  
1302 in these hypothalamic nucleus, (e.g. paraventricular nucleus, which are involved in the  
1303 blood pressure regulation), may leads to long-term cardiovascular disease [15, 28].

1304         Our data shows that HFD in adolescence leads rats to present an increase in SBP  
1305 and MBP at adulthood, corroborating with studies of metabolic programming by HFD  
1306 maternal exposure [15, 29]. Studies outside metabolic programming context, have widely  
1307 discussed that cardiovascular function is controlled by diverse mechanisms, mainly  
1308 sympathetic and parasympathetic activity that innervate the blood vessels and heart. After  
1309 hexamethonium exposure, an expressive decrease in the DBP, SBP and MBP was  
1310 observed in HFD group compared to control group. Thus, the increase in BP showed by  
1311 HFD animals in later life could be due to a programming effect of HFD during pubertal  
1312 period that lead to sympathetic nervous system (SNS) dysfunction. In addition, the higher  
1313 sensitivity to ganglionic blockade of hexamethonium chloride, and consequent expressive  
1314 decrease in BP, strongly demonstrate that increased BP observed here support this idea.  
1315 Some reports have shown that male Wistar rats shown higher MBP as well as renal  
1316 sympathetic nerve activity in HFD rats compare to control [30]. In addition, rabbits fed  
1317 HFD, for only four weeks, also shown higher BP combined with increase in renal  
1318 sympathetic nerve activity [31]. Although both studies above are outside metabolic  
1319 programming concept, taken together, these data suggest that HFD exposure has a major  
1320 impact on development of CNS during adolescence, like as other previous period of life  
1321 [26, 27, 32].

1322         Besides the impairment of HFD exposure on control of BP, in this study HFD  
1323 exposure during adolescence leads to dyslipidemia with increase in triglyceride levels at  
1324 adulthood. Although no difference was observed in total cholesterol between de groups,  
1325 similarly was found in other programming window [29, 33]. Triglyceride (TGL) is  
1326 considered as an important biomarker of cardiovascular risk and this increase due to HFD  
1327 exposure is well discussed in a lot of studies [34, 35]. In addition, no difference was  
1328 observed in fasting glucose, HFD exposure is often associated to glucose intolerance [9,  
1329 17]. Accordingly, in our study HFD treatment in adolescent rats showed to be able on  
1330 increased body weight gain associated to glucose intolerance at adulthood, corroborating  
1331 with previous studies [17, 23, 36]. Taken together, our data appear to reveal that dietary

1332 insult during adolescence might lead to programming to increase risk factors of  
1333 cardiometabolic syndrome at adulthood and emphasized peripubertal period as a crucial  
1334 window of development, as well as on gestation and lactation [9, 11, 37].

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1356 **5. Conclusion**

1357           In summary, we demonstrate that HFD consumption during adolescence is able to  
1358 program cardiometabolic dysfunctions in male rats, and in this sense is strongly  
1359 associated to cardiovascular risk, leading rats to dyslipidemia, mainly  
1360 hypertriglyceridemia, obesity, glucose intolerance and hypertension at adulthood. The  
1361 present study confirms the susceptibility of adolescence as a critical window of  
1362 development, pointing to the need of intervention and control of access to HFD in this  
1363 life period to prevent cardiometabolic disease.

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1381 **Declaration of interest**

1382 The authors declare no conflict of interest associated with this manuscript.

1383

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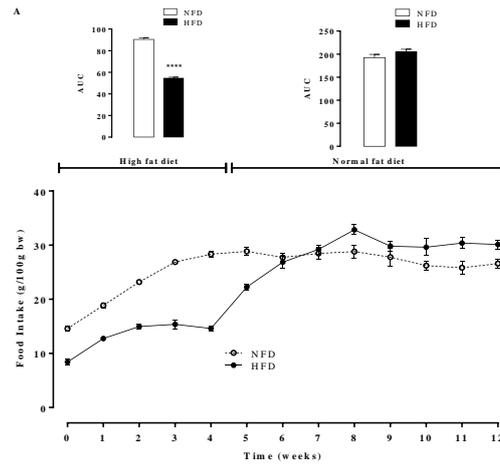
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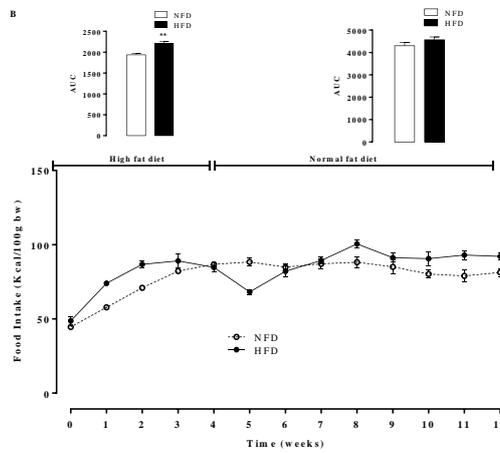
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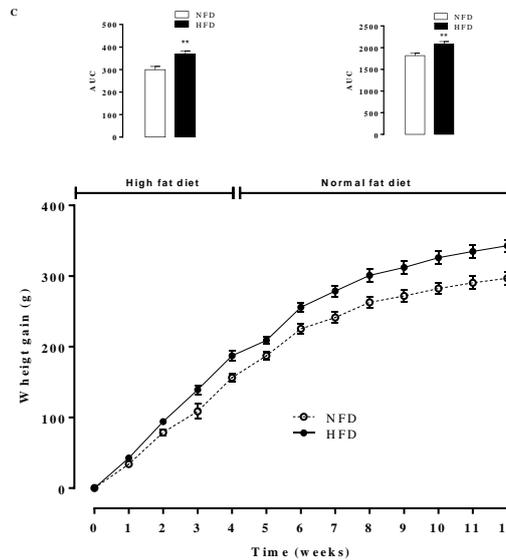
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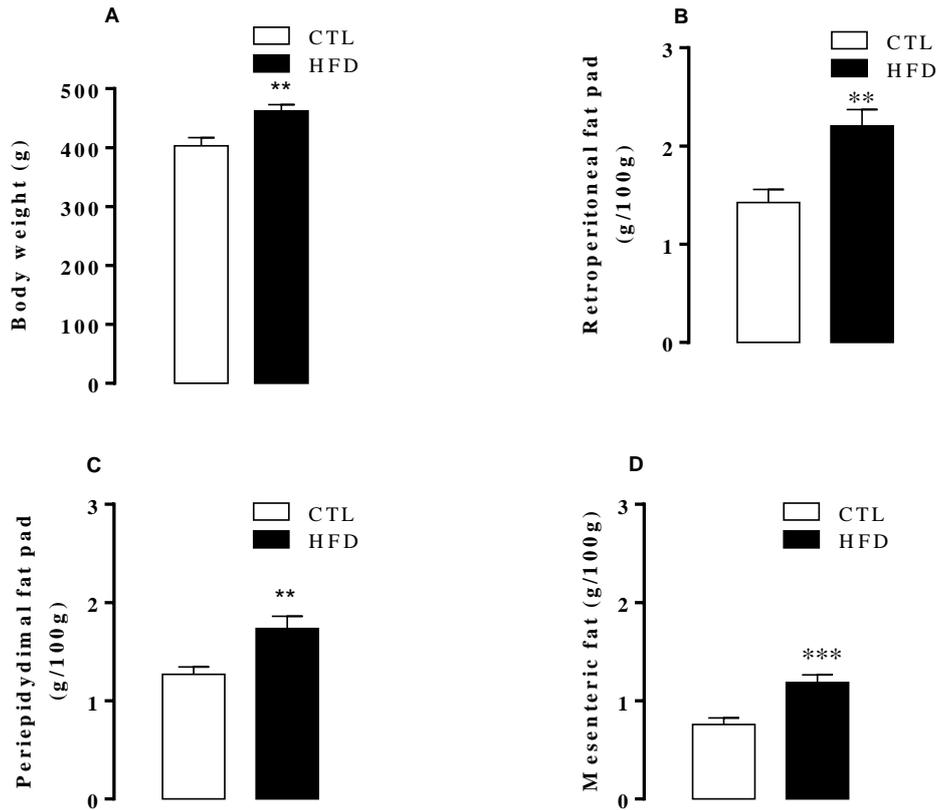
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1535 **Fig 1:** Food intake relative to 100g of body weight and its total area-under-the-curve (AUC) (A),  
1536 Food intake relative to kcal/100g of body weight and its AUC (B), body weight gain and its AUC  
1537 (C) from 30 to 120 days of life from animals exposed to normal diet (NFD, white) and high-fat-  
1538 diet (HFD, black). n = 20 animals; values are mean  $\pm$  SEM; \*\*\*\*p<0.0001, \*\*p<0.01 indicate  
1539 statistical significance by Student's t-test.



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1541 **Fig 2:** Body weight (A), retroperitoneal (B), periepididymal (C), and mesenteric fat stores (D)  
 1542 from 120-days-old rats exposed to normal fat diet (NFD, white) and high-fat-diet (HFD, black)  
 1543 at adolescence. n = 10 animals; values are mean ± SEM; \*\*\*p<0.001, \*\*p<0.01 indicate statistical  
 1544 significance by Student's t-test.

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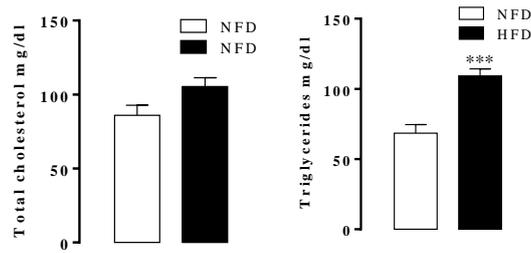
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1558 **Fig 3:** Total cholesterol (A) and triglycerides (B), from 120-days-old rats exposed to normal fat  
1559 diet (NFD, white) and high-fat-diet (HFD, black). n = 6 animals for total cholesterol and n = 6-8  
1560 for triglycerides; values are mean  $\pm$  SEM; \*\*\*p<0.001 indicate statistical significance by  
1561 Student's t-test.

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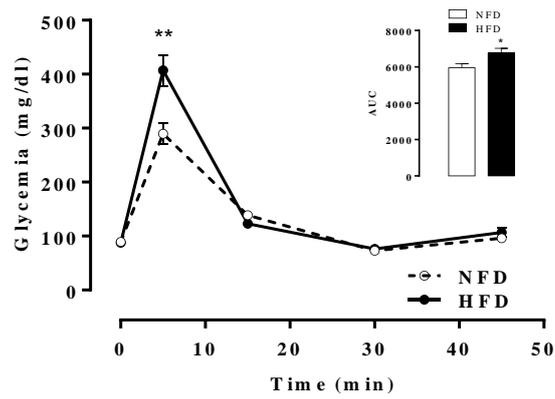
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1580 **Fig 4:** Glucose tolerance test and its total area-under-the-curve (AUC) from 120-days-old rats  
 1581 exposed to normal fat diet (NFD, white) and high-fat-diet (HFD, black). n = 6-9 animals for test;  
 1582 values are mean ± SEM; \*\*p<0.001, \*p<0.05 indicate statistical significance by Student's t-test.

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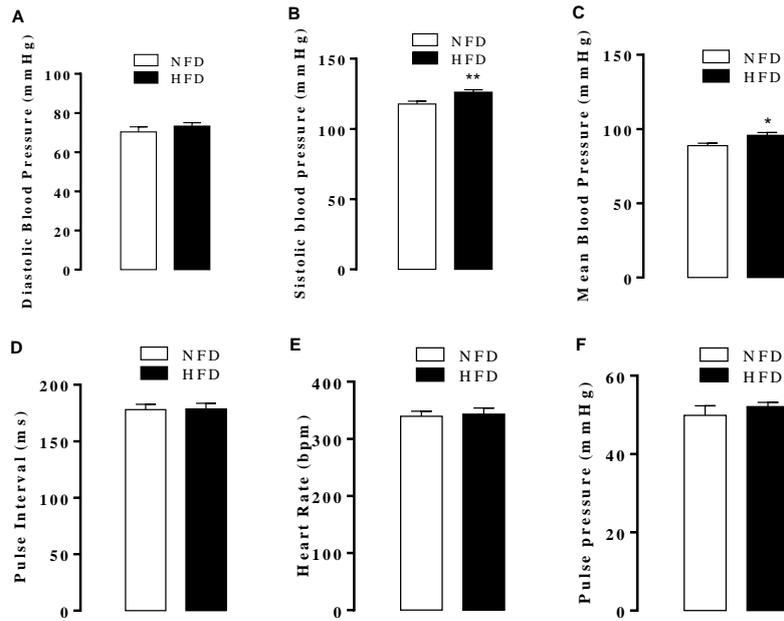
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1594 **Fig 5:** Diastolic blood pressure (DBP; A), systolic blood pressure (SBP; B), mean blood pressure  
 1595 (MBP; C), pulse interval (PI; D), heart rate (HR; E) and pulse pressure (PP; F) from 120-days-  
 1596 old rats exposed to normal fat diet (NFD, white; n = 11) and high-fat-diet (HFD, black; n = 13).  
 1597 Values are mean ± SEM; \*\*p<0.001 and \*p<0.05 indicate statistical significance by Student's t-  
 1598 test.

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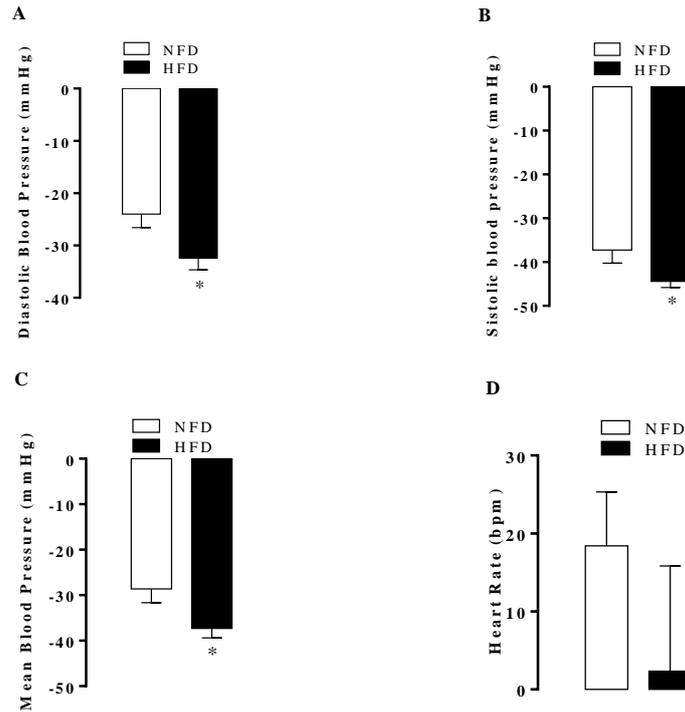
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1609 **Fig 6:** Depressant response to hexamethonium in diastolic blood pressure (DBP; A), systolic  
 1610 blood pressure (SBP; B), mean blood pressure (MBP; C), and heart rate (HR; D) in 120–days-old  
 1611 rats exposed to normal fat diet (NFD, white; n = 11) and high-fat-diet (HFD, black; n = 13).  
 1612 Values are mean ± SEM; \*p<0.05 indicate statistical significance by Student’s t-test.

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